

University of Dundee

Mathematical Modelling of Cancer Invasion

Bitsouni, Vasiliki; Chaplain, Mark; Eftimie, Raluca

Published in:
Mathematical Models and Methods in Applied Sciences

DOI:
[10.1142/S021820251750035X](https://doi.org/10.1142/S021820251750035X)

Publication date:
2017

Licence:
CC BY

Document Version
Publisher's PDF, also known as Version of record

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):
Bitsouni, V., Chaplain, M., & Eftimie, R. (2017). Mathematical Modelling of Cancer Invasion: The Multiple Roles of TGF- β Pathway on Tumour Proliferation and Cell Adhesion. *Mathematical Models and Methods in Applied Sciences*, 27(10), 1929-1962. <https://doi.org/10.1142/S021820251750035X>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Mathematical modelling of cancer invasion: The multiple roles of TGF- β pathway on tumour proliferation and cell adhesion

Vasiliki Bitsouni

*Division of Mathematics, University of Dundee,
Dundee, DD1 4HN, Scotland, UK
vbitsouni@maths.dundee.ac.uk
vbitsouni@gmail.com*

Mark A. J. Chaplain

*School of Mathematics and Statistics,
Mathematical Institute (MI), North Haugh,
University of St Andrews, St Andrews, KY16 9SS,
Scotland, UK
majc@st-andrews.ac.uk*

Raluca Eftimie

*Division of Mathematics, University of Dundee,
Dundee, DD1 4HN, Scotland, UK
reftimie@maths.dundee.ac.uk*

Received 9 March 2017

Revised 25 April 2017

Accepted 14 May 2017

Published 6 July 2017

Communicated by N. Bellomo

In this paper, we develop a non-local mathematical model describing cancer cell invasion and movement as a result of integrin-controlled cell–cell adhesion and cell–matrix adhesion, and transforming growth factor-beta (TGF- β) effect on cell proliferation and adhesion, for two cancer cell populations with different levels of mutation. The model consists of partial integro-differential equations describing the dynamics of two cancer cell populations, coupled with ordinary differential equations describing the extracellular matrix (ECM) degradation and the production and decay of integrins, and with a parabolic PDE governing the evolution of TGF- β concentration. We prove the global

This is an Open Access article published by World Scientific Publishing Company. It is distributed under the terms of the Creative Commons Attribution 4.0 (CC-BY) License. Further distribution of this work is permitted, provided the original work is properly cited.

existence of weak solutions to the model. We then use our model to explore numerically the role of TGF- β in cell aggregation and movement.

Keywords: Non-local model of cancer progression; existence; boundedness of solution; cell heterogeneity; TGF- β ; cell-cell and cell-matrix adhesion.

AMS Subject Classification: 35A01, 35Q92, 35R09, 92C15, 92C17, 92-08

1. Introduction

Cellular adhesion, i.e. cell-cell and cell-matrix adhesion, and cellular proliferation are fundamental features of multicellular organisms, linked to maintenance of order in the organisms, e.g. tissue formation, stability and breakdown.³ These interactions between cells and the extracellular matrix (ECM) are mediated through cell surface receptors, a major group of which is represented by the integrins,⁶⁸ and various cytokines and chemokines. Another group of molecules involved in cell-cell adhesion is represented by the cadherin families.²⁶ There are several signalling pathways that control normal cell processes like cell proliferation, division, cellular adhesion and apoptosis, with transforming growth factor β (TGF- β) pathway to be one of the most critical. Belonging to a large family of multifunctional polypeptides, TGF- β regulates the proliferation, differentiation, adhesion, migration and apoptosis of many cell types, including endothelial cells, hematopoietic cells and lymphocytes,⁴⁷ and ECM production.³¹

Various signals, including integrin, Notch, Wnt, TNF- α , and EGF signals, have been reported to cooperate or synergize with TGF- β signalling and stimulate tumour invasion and metastasis.⁴⁷ Experimental studies⁴⁴ showed that the loss of TGF- β responsiveness is one of the events that initiate fibrotic disease and malignant progression of cancer, as well as cancer metastasis.⁶⁵ TGF- β induces morphological, biochemical and transcriptional changes towards a mesenchymal phenotype, a process called epithelial to mesenchymal transition (EMT) (see Refs. 42 and 51 and many references therein). EMT occurs when epithelial cells lose their epithelial cell characteristics and become mesenchymal. Mesenchymal cells can return to an epithelial phenotype, a process called mesenchymal-epithelial transition (MET). Through these processes, cancer cells become metastatic and form new colonies at distant sites.

Experimental studies^{33,41} have shown that tumours consist of heterogeneous populations of cells, which are the result of genetic instability. Intra-tumour heterogeneity appears in almost all phenotypic cell features: from cell morphology, to gene expression, motility, proliferation, immunogenicity and metastatic potential.⁴⁵ While both normal cells and cancer cells appear to be heterogeneous for various characteristics (e.g. surface antigens), cellular heterogeneity is shown⁵⁴ to be more pronounced in malignant neoplasms. Experimental studies have shown complex interactions between clonal cancer cell sub-populations in heterogeneous tumours: from stable coexistence to competitive exclusion.³⁸ The metastatic and invasive potential of heterogeneous tumours is influenced by the interactions among the

cells, and the interactions between cells and ECM components. To detach from the main aggregation/tissue, cells loose cell-cell adhesion and strengthen cell-matrix adhesion (these changes in cell-cell/cell-ECM adhesion can be influenced by TGF- β signalling), which leads to ECM remodelling and degradation (with the help of enzymes called matrix metalloproteinases; MMPs).

Over the last three decades there have been multiple mathematical models introduced to investigate the formation and movement of various cell aggregations (see, for example, Refs. 1, 3, 5, 6, 8, 48, 49, 55, 56 and 59, and the many references therein). While there are mathematical models in the literature that investigate the roles of TGF- β on cancer dynamics, generally these models focus on particular aspects of cancer progression (e.g. growth^{46,63}). There are very few models that investigate, in an integrated manner, the multiple roles of TGF- β on cancer evolution (see, for example Refs. 4 and 67), and in general these models focus on the motility and growth rate of early stage cancer cells.

In this study, we present a novel mathematical model which investigates in a integrated manner the various roles of TGF- β on tumour growth/decay, and on cell-cell and cell-matrix interactions, but paying particular attention to the opposite role of TGF- β on early stage versus late stage cancer cells. To this end, we introduce a non-local hyperbolic-parabolic model for cell-cell and cell-matrix adhesion for two cancer cell populations: an early stage cancer population, moving both randomly and in a directed manner in response to cell-cell and cell-matrix adhesive forces, and a late stage cancer population (i.e. a mutated clone) moving only in a directed manner following self-adhesive and cross-adhesive cell-cell forces, as well as matrix interactions. Since TGF- β does not affect only tumour growth, but impacts also cell adhesion,⁴ we model the interactions between TGF- β and integrins that influence the cell-cell and cell-matrix adhesive forces. The computational results show a range of heterogeneous invasion patterns, as a result of the opposite role of TGF- β in early and late stages of cancer. Analytical results show the global existence of bounded solutions (hence existence of various types of invasion patterns). We note that existence results have been shown for local nonlinear parabolic PDEs for cell movement coupled with ODEs describing the ECM dynamics with tissue remodelling,^{22,32,61,62} as well as for non-local parabolic models describing cancer invasion when the ECM production is zero¹⁰ and when it is nonzero.⁶⁰ Existence results have also been shown for local hyperbolic models for chemo-sensitive movement.²⁹ However, in contrast to these previous results, here we show existence for a non-local parabolic-hyperbolic model for cancer cell movement.

The structure of this paper is as follows. In Sec. 2, we present our mathematical model, which consists of partial integro-differential equations describing the dynamics of cell populations in early and late stages of cancer, coupled with ordinary differential equations describing ECM and integrins dynamics, and a parabolic partial differential equation describing TGF- β dynamics. In Sec. 3, we present a suitable notion of weak solution to the model and we prove the global-in-time existence

of bounded solutions to our system as the vanishing viscosity limit of a classical solution to an associated parabolic problem. In Sec. 4, we undertake numerical simulations to investigate the effect of TGF- β on cancer cell movement, and observe a range of patterns obtained for different values of parameters of the model. Finally, in Sec. 5, we summarise our results and give some concluding remarks.

2. The Mathematical Model of TGF- β Regulatory Network in Cancer

TGF- β plays a crucial role in embryonic development, wound healing and cancer. Moreover, TGF- β signalling stimulates EMT in certain epithelial cells (see Ref. 51 and many references therein) and consequently induces various diseases, including cancer. The way that TGF- β interacts with cancer cells varies between early and late stages of cancer (see Fig. 1), making its behaviour difficult to analyse. We consider a two-population model describing the behaviour of an early stage cancer population and a late stage cancer (descendant clone) population, which interact with each other, as well as with the ECM, via long-range integrin-controlled adhesive and repulsive forces^{18,23} on bounded spatial domain $\Omega \subset \mathbb{R}^n$ with smooth boundary $\partial\Omega$. For $T > 0$ let $\Omega_T = (0, T) \times \Omega$. Denote by $u_1(t, x)$ and $u_2(t, x)$ the density of early and late stage cancer cells, respectively, at position x and time t , by $f(t, x)$ the ECM density, and by $c(t, x)$ the density of integrin receptors on the surface of cancer cells (receptors involved in cell-cell and cell-matrix interactions). Finally, we denote by $b(t, x)$ the TGF- β concentration. For compact notation, we define the vectors $\underline{u}(t, x) = (u_1(t, x), u_2(t, x))^T$ and $\underline{v}(t, x) = (\underline{u}(t, x), f(t, x))^T$.

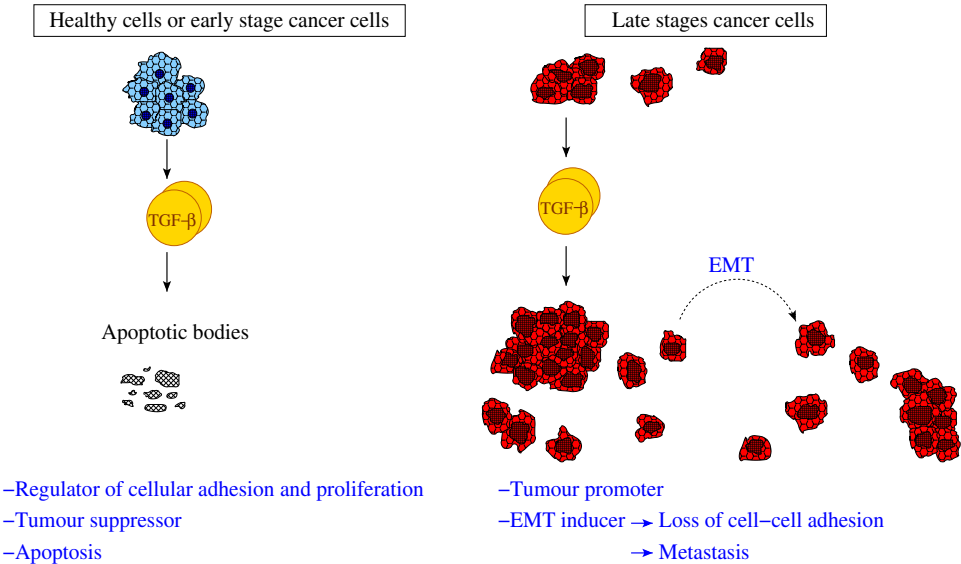


Fig. 1. A caricature summarising the dual role of TGF- β in cancer progression.

Cancer cells dynamics. Cancer cells can switch from a homogeneous type of invasion to a heterogeneous type of invasion described by (directionally moving) invading chains.¹² Therefore, we assume that the movement of the early stage cancer cell population u_1 is governed by random motility (which underlines a homogeneous type of invasion), as well as directed motility in response to cell-cell and cell-matrix adhesive forces (which underlines the heterogeneous type of invasion).⁹ Let D_u describe the random motility coefficient and $F_1[\underline{u}, f, c, b]$ describe the non-local directed motility. In contrast, the late stage cancer cell population, u_2 , moves only in a directed manner (hence exhibiting a heterogeneous type of invasion) in response to cell-cell and cell-matrix adhesion forces (described by a non-local term $F_2[\underline{u}, f, c, b]$). Moreover, the u_1 cells can mutate into u_2 cells at a constant rate M . TGF- β has been found to have bidirectional functions in the progression of cancer. In early stages of cancer, TGF- β is an antiproliferative and proapoptotic signal, while in late stages of cancer it acts as a tumour promoter.⁵⁷ Thus, we have the following equations describing the dynamics of the two cancer cell populations:

$$\frac{\partial u_1}{\partial t} = D_u \Delta u_1 - \nabla \cdot (u_1 F_1[\underline{u}, f, c, b]) - M u_1 + G_1(\underline{u}, b), \quad (2.1a)$$

$$\frac{\partial u_2}{\partial t} = -\nabla \cdot (u_2 F_2[\underline{u}, f, c, b]) + M u_1 + G_2(\underline{u}, b). \quad (2.1b)$$

Taking into account the effect of TGF- β on cancer cell proliferation and assuming that both u_1 and u_2 cells can proliferate in a logistic manner (to describe the observed slow-down in tumour growth following the loss of nutrients³⁷), we choose the growth functions to be given by

$$G_i(\underline{u}, b) = r_i u_i \left(1 - \frac{u_1 + u_2}{k_u} \right) \left(1 + (-1)^i c_b \frac{b}{b_m} \right), \quad i = 1, 2, \quad (2.2)$$

where r_1 and r_2 are the growth rates of the u_1 and u_2 populations, respectively, k_u is the carrying capacity, b_m is the maximum TGF- β concentration, and c_b is a coefficient related to the effect of TGF- β on cancer cell proliferation/decay. In particular, the term $(-1)^i$ models the anti-tumour effect of TGF- β on early tumours ($i = 1$), and the pro-tumour effect on late tumours ($i = 2$). Note that these growth functions incorporate also the principle of competition between clonal sub-populations in heterogeneous tumours.³⁸

The non-local cell-cell and cell-matrix adhesion and repulsion forces for cancer cell populations u_1 and u_2 , are described by a function that depends on cell densities, ECM and integrin densities, and concentrations of TGF- β molecules

$$F_i : C(\bar{\Omega} : \mathbb{R})^5 \mapsto C^{1,\zeta}(\bar{\Omega} : \mathbb{R}^n), \quad \zeta \in (0, 1], \quad i = 1, 2, \quad (2.3)$$

given by the following relation

$$F_i[\underline{u}, f, c, b](x) := \int_{\Omega} K(|y - x|) g_i(\underline{u}(y), f(y), c(x), b(x)) dy, \quad i = 1, 2, \quad (2.4)$$

where $K \in L^\infty(\Omega)$, with $\partial_x K \in L^\infty(\Omega)$. The functions $g_i(\underline{u}, f, c, b)$, $i = 1, 2$, describe the nature of the cell-cell and cell-matrix adhesive forces. These functions increase

when the cell density and ECM density increase, and accordingly they decrease when the cell density and ECM density decrease. The functions $g_i, i = 1, 2$, are given by

$$g_i(\underline{u}, f, c, b) := S_i(c, b)u_i + S(c, b)u_j + C_i(c, b)f, \quad i, j = 1, 2, i \neq j, \quad (2.5)$$

where $S_i(c, b)$ is the cell-cell self-adhesion strength function for populations u_i , $S(c, b)$ is the cell-cell cross-adhesion strength function between the two populations, and $C_i(c, b)$ is the adhesion strength function between population u_i and ECM.

Integrins are molecules known to have a regulative role in cell-cell and cell-matrix adhesion,⁴ while the role of TGF- β in cellular adhesion is dual: (i) Promotes cell-matrix adhesion by inducing the synthesis and the secretion of ECM-adhesion molecules laminin and fibronectin and the upregulation of integrin expression for these matrix-adhesion molecules^{30,66}; (ii) Decreases cell-cell adhesion.^{52,67} Thus, to define these adhesion strength functions we consider the integrin density, c , and TGF- β concentration, b . Since cell mutation could lead to more integrins,³⁵ we consider strength functions with different integrin levels for each of the two populations. The more integrins a cell has, the stronger its adhesion force. Therefore, biologically realistic choices for these adhesion strength functions are the increasing, bounded and positive functions given by:

$$\begin{aligned} S_i(c, b) &= s_i^*(1 + \tanh(a_i c - a_{b_i} b)), \quad S(c, b) = s^*(1 + \tanh(dc - d_b b)), \\ C_i(c, b) &= c_i^*(1 + \tanh(e_i c + e_{b_i} b)), \quad i = 1, 2, \end{aligned} \quad (2.6)$$

where $a_i, a_{b_i}, d, d_b, e_i, e_{b_i}$ and $s_i^*, s^*, c_i^*, i = 1, 2$, are positive real numbers.

ECM dynamics. The ECM is considered as non-motile matter, with changes to its density due to degradation by u_1 and u_2 cell populations upon contact at rates $\alpha > 0$ and $\beta > 0$, respectively, and ECM density remodelling back to normal levels, at a constant rate of $\delta > 0$. Moreover, TGF- β induces the synthesis of ECM adhesion molecules,^{66,67} at a rate of $\theta_\beta > 0$. Thus the dynamics of ECM, $f(t, x)$, is described by:

$$\frac{\partial f}{\partial t} = -\alpha u_1 f - \beta u_2 f + \theta_\beta b f + \delta f \left(1 - \frac{f}{f_m}\right), \quad (2.7)$$

where f_m is the maximum ECM density at which the ECM fills up all available physical space.

Integrin dynamics. We assume that the level of integrins depends on cancer cell density, such that cell mutation changes the density of receptors (since in highly metastatic cancers, the expression of integrins is up-regulated³⁵). Moreover, TGF- β signalling up-regulates the integrin expression,^{4,42} at a rate of p_3 . Therefore, the dynamics of integrins, $c(t, x)$, can be described by:

$$\frac{\partial c}{\partial t} = p_1 u_1 + p_2 u_2 + p_3 b c - q c, \quad (2.8)$$

where q is the decay rate of c , and p_1 and p_2 are the production rates of integrins by u_1 and u_2 cancer cell populations, respectively. To model the increase in receptors on highly mutated cancer cells, we assume that $p_2 > p_1$ (see Table A.2).

TGF- β dynamics. Finally, TGF- β is assumed to diffuse freely in the spatial domain, after being released by u_1 and u_2 cells, and decay at a rate of $q_b > 0$. Therefore, the dynamics of TGF- β , $b(t, x)$, is described by:

$$\frac{\partial b}{\partial t} = D_b \Delta b + \lambda(\underline{u}) - q_b b, \quad (2.9)$$

where D_b is the TGF- β diffusion coefficient and $\lambda(\underline{u})$ is the TGF- β production term. Here, we choose $\lambda(\underline{u}) = \mu_1 u_1 + \mu_2 u_2$, with μ_1 and μ_2 to be the production rates of TGF- β by u_1 and u_2 , respectively.

The relations (2.1)–(2.9) are summarised in the following system:

$$\frac{\partial u_1}{\partial t} = D_u \Delta u_1 - \nabla \cdot (u_1 F_1[\underline{u}, f, c, b]) - M u_1 + G_1(\underline{u}, b), \quad (2.10a)$$

$$\frac{\partial u_2}{\partial t} = -\nabla \cdot (u_2 F_2[\underline{u}, f, c, b]) + M u_1 + G_2(\underline{u}, b), \quad (2.10b)$$

$$\frac{\partial f}{\partial t} = -\alpha u_1 f - \beta u_2 f + \theta_\beta b f + \delta f \left(1 - \frac{f}{f_m}\right), \quad (2.10c)$$

$$\frac{\partial c}{\partial t} = p_1 u_1 + p_2 u_2 + p_3 b c - q c, \quad (2.10d)$$

$$\frac{\partial b}{\partial t} = D_b \Delta b + \mu_1 u_1 + \mu_2 u_2 - q_b b. \quad (2.10e)$$

We impose the following initial conditions:

$$\begin{aligned} u_i(0, x) = u_{i0}(x) \geq 0, \quad i = 1, 2, \quad f(0, x) = f_0(x) \geq 0, \\ c(0, x) = c_0(x) \geq 0, \quad b(0, x) = b_0(x) \geq 0, \quad \text{in } \Omega. \end{aligned} \quad (2.11)$$

Finally, we assume that there is no-flux of both cancer cells and TGF- β proteins on the boundary of the domain,

$$\langle \nabla u_i, \nu \rangle = 0 = \langle \nabla b, \nu \rangle, \quad i = 1, 2, \quad \text{on } (0, \infty) \times \partial\Omega \quad (2.12)$$

and

$$\langle F_i[\underline{u}, f, c, b], \nu \rangle = 0, \quad i = 1, 2, \quad \text{on } (0, \infty) \times \partial\Omega, \quad (2.13)$$

where ν is the outward unit normal vector to $\partial\Omega$.

3. Existence of Solution

To prove the existence of solution for system (2.10), we use the theory of semi-groups combined with the vanishing viscosity method (to transform Eq. (2.10b) into a parabolic equation). Then we show that in the vanishing viscosity limit, we obtain weak solutions for (2.10). We note that the steps in this proof of existence

of approximate solution follow similar approaches taken in Ref. 10, where a simpler parabolic-ODE model with no production term in the ODE (i.e. the ODE in Ref. 10 contains only decay term, which implies global boundedness of its solution) is considered, whereas in our model the production terms in (2.10c)–(2.10d) add an extra layer of complexity to the proof. Moreover, for the proof of vanishing viscosity limit we use techniques similar to those in Ref. 29 for a hyperbolic–elliptic model. The link between these two proofs is based on the extraction of the appropriate estimates for the vanishing viscosity limit, for our more complex system of non-local parabolic–hyperbolic equations coupled with ODEs.

3.1. Existence of approximate solution

We will approximate system (2.10) by the following system:

$$\begin{aligned} \frac{\partial u_1^\epsilon}{\partial t} - D_u \Delta u_1^\epsilon + M u_1^\epsilon &= -\nabla u_1^\epsilon \cdot F_1[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] \\ &\quad - u_1^\epsilon (\nabla \cdot F_1[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]) + G_1(u_1^\epsilon, u_2^\epsilon, b^\epsilon), \end{aligned} \quad (3.1a)$$

$$\begin{aligned} \frac{\partial u_2^\epsilon}{\partial t} - \epsilon \Delta u_2^\epsilon + u_2^\epsilon &= -\nabla u_2^\epsilon \cdot F_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] \\ &\quad - u_2^\epsilon (\nabla \cdot F_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]) + h_1(u_1^\epsilon, u_2^\epsilon, b^\epsilon), \end{aligned} \quad (3.1b)$$

$$\frac{\partial f^\epsilon}{\partial t} = h_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon) f^\epsilon - \frac{\delta}{f_m} f^{\epsilon^2}, \quad (3.1c)$$

$$\frac{\partial c^\epsilon}{\partial t} = h_3(u_1^\epsilon, u_2^\epsilon) + c^\epsilon h_4(b^\epsilon), \quad (3.1d)$$

$$\frac{\partial b^\epsilon}{\partial t} - D_b \Delta b^\epsilon + q_b b^\epsilon = h_5(u_1^\epsilon, u_2^\epsilon), \quad (3.1e)$$

for $0 < \epsilon \leq 1$, where

$$\begin{aligned} h_1(u_1^\epsilon, u_2^\epsilon, b^\epsilon) &= M u_1^\epsilon + u_2^\epsilon + G_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon), \\ h_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon) &= -\alpha u_1^\epsilon - \beta u_2^\epsilon + \theta_\beta b^\epsilon + \delta, \\ h_3(u_1^\epsilon, u_2^\epsilon) &= p_1 u_1^\epsilon + p_2 u_2^\epsilon, \quad h_4(b^\epsilon) = p_3 b^\epsilon - q \quad \text{and} \\ h_5(u_1^\epsilon, u_2^\epsilon) &= \mu_1 u_1^\epsilon + \mu_2 u_2^\epsilon. \end{aligned} \quad (3.2)$$

The ICs are given by

$$\begin{aligned} u_i^\epsilon(0, x) &= u_{i_0}(x) \geq 0, \quad i = 1, 2, \quad f^\epsilon(0, x) = f_0(x) \geq 0, \\ c^\epsilon(0, x) &= c_0(x) \geq 0, \quad b^\epsilon(0, x) = b_0(x) \geq 0, \quad \text{in } \Omega. \end{aligned} \quad (3.3)$$

Finally, the BCs corresponding to (2.12)–(2.13) are given by the relations

$$\langle \nabla u_i^\epsilon, \nu \rangle = 0 = \langle \nabla b^\epsilon, \nu \rangle, \quad i = 1, 2, \quad \text{on } (0, \infty) \times \partial\Omega \quad (3.4)$$

and

$$\langle F_i[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon], \nu \rangle = 0, \quad i = 1, 2, \quad \text{on } (0, \infty) \times \partial\Omega. \quad (3.5)$$

For the full non-local interaction terms (2.3)–(2.4) we make the following assumptions:

$$F_i[u_1, u_2, f, c, b](x) = \int_{\Omega} N_i(x, y, u_1(y), u_2(y), f(y), c(x), b(x)) dy, \quad (3.6)$$

where $N_i : \Omega^2 \times \mathbb{R}^5 \mapsto \mathbb{R}^n$, $i = 1, 2$, is a continuous function, which satisfies

$$N_i(x, y, \underline{0}, c, b) = 0, \quad i = 1, 2, \quad \text{for all } (x, y) \in \Omega^2, \quad c, b \in \mathbb{R}, \quad (3.7)$$

and

$$N_i(\cdot, y, \phi, \chi, \psi, c, b) \in C^{1, \zeta}(\bar{\Omega} : \mathbb{R}^n), \quad i = 1, 2, \quad \text{for all } y \in \Omega, \quad (\phi, \chi, \psi) \in \mathbb{R}^3. \quad (3.8)$$

Since functions $S_i(c, b)$, $S(c, b)$, $C_i(c, b)$, $i = 1, 2$, (given by (2.6)) are bounded, we assume that there is a constant L_N , which depends on the bound for S_i, S, C_i , $i = 1, 2$, such that for any $\phi_1, \phi_2, \chi_1, \chi_2, \psi_1, \psi_2 \in \mathbb{R}$ we have

$$\begin{aligned} & |N_i(x, y, \phi_1, \chi_1, \psi_1, c, b) - N_i(x, y, \phi_2, \chi_2, \psi_2, c, b)| \\ & + |\partial_x N_i(x, y, \phi_1, \chi_1, \psi_1, c, b) - \partial_x N_i(x, y, \phi_2, \chi_2, \psi_2, c, b)| \\ & \leq L_N(|\phi_1 - \phi_2| + |\chi_1 - \chi_2| + |\psi_1 - \psi_2|), \quad i = 1, 2, \end{aligned} \quad (3.9)$$

uniformly with respect to $(x, y) \in \Omega^2$.

We assume that h'_2, h'_3 and h'_4 exist, and that

$$h'_2 : \mathbb{R} \times \mathbb{R} \times \mathbb{R} \mapsto \mathbb{R}, \quad h'_3 : \mathbb{R} \times \mathbb{R} \mapsto \mathbb{R} \quad \text{and} \quad h'_4 : \mathbb{R} \mapsto \mathbb{R} \quad (3.10)$$

are locally Lipschitz functions.

Based on relation (2.2) we can assume that there are constants $B_i, D_i > 0$, $i = 1, 2$, such that for $u_1^\epsilon, u_2^\epsilon, b^\epsilon \geq 0$, $i = 1, 2$, we have

$$G_i(u_1^\epsilon, u_2^\epsilon, b^\epsilon) \leq B_i - D_i u_i^\epsilon, \quad i = 1, 2. \quad (3.11)$$

Moreover, based on relation (3.2) we can assume that there are constants $\Lambda_j > 0$, $j = 1, \dots, 4$, such that for $u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon \geq 0$:

$$f^\epsilon h_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon) - \frac{\delta}{f_m} f^{\epsilon^2} \leq \Lambda_1 b^\epsilon - \Lambda_2 f^\epsilon \quad (3.12)$$

and

$$c^\epsilon h_4(b^\epsilon) \leq \Lambda_3 b^\epsilon - \Lambda_4 c^\epsilon. \quad (3.13)$$

We now consider the sectorial operators

$$A_1 = -D_u \Delta + MI, \quad A_2 = -\epsilon \Delta + I \quad \text{and} \quad A_3 = -D_b \Delta + q_b I \quad (3.14)$$

in the space $X = L^p(\Omega)$, with common domain of definition

$$D = D(A_1) = D(A_2) = D(A_3) = \left\{ w \in W^{2,p} : \frac{\partial w}{\partial \nu} = 0 \text{ on } \partial\Omega \right\}, \quad (3.15)$$

and $\operatorname{Re}(\sigma(A_j)) > 0$, $j = 1, 2, 3$. Then the fractional powers are well defined

$$X^\gamma = D(A_1^\gamma) = D(A_2^\gamma) = D(A_3^\gamma), \quad 0 < \gamma < 1, \quad (3.16)$$

with the graph norm

$$\|x\|_{X^\gamma} = \|A_j^\gamma x\|_X, \quad \text{for } x \in X^\gamma, \quad j = 1, 2, 3. \quad (3.17)$$

Then from Ref. 27, we have the following embeddings:

$$X^\gamma \subset W^{1,p}(\Omega) \quad \text{for } \gamma > \frac{1}{2}, \quad (3.18)$$

$$X^\gamma \subset C^{0,r}(\bar{\Omega}) \quad \text{for } \frac{r}{2} + \frac{n}{2p} < \gamma < \frac{1}{2} + \frac{n}{2p}, \quad 0 < r < 1, \quad (3.19)$$

where $C^{0,r}(\bar{\Omega})$ is the space of all Hölder continuous functions with exponent r in Ω . Notice that for

$$\gamma \in \left(\frac{1}{2}, \frac{1}{2} + \frac{1}{2p}\right) \quad \text{for } p > n, \quad (3.20)$$

(3.18) and (3.19) are satisfied.

Moreover, since A_1, A_2 and A_3 are sectorial operators, then each of $-A_1, -A_2$ and $-A_3$ is the infinitesimal generator of an analytic semigroup $\{e^{-tA_j}\}_{t \geq 0}$, $j = 1, 2, 3$. Therefore, there exists a positive constant C_γ such that the following inequality holds²⁷:

$$\|A_j^\gamma e^{-A_j t} w\|_X \leq C_\gamma t^{-\gamma} e^{-\xi_j t} \|w\|_X, \quad \text{for } w \in X, \quad (3.21)$$

where $0 < \xi_j < \operatorname{Re}(\sigma(A_j))$, $j = 1, 2, 3$, and

$$\|A_j^\gamma e^{-A_j t} w\|_X \leq k_\gamma \|w\|_{X^\gamma}, \quad \text{for } w \in X^\gamma, \quad (3.22)$$

where k_γ positive constant.

Theorem 3.1. *Let $u_1^\epsilon(0, x), u_2^\epsilon(0, x), b^\epsilon(0, x) \in X^\gamma$ and $f^\epsilon(0, x), c^\epsilon(0, x) \in W^{1,p}(\Omega)$. If assumptions (3.3)–(3.13) and (3.20) are satisfied, then for any $T > 0$ there exists a unique global-in-time solution $(u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon) \in C([0, T]; [X^\gamma]^2 \times [W^{1,p}(\Omega)]^2 \times X^\gamma)$ to (3.1)–(3.5), which remains bounded and the bounds are ϵ -independent. Moreover, the solution satisfies*

$$(u_1^\epsilon, u_2^\epsilon, b^\epsilon) \in C^1((0, T); [X^\gamma]^3) \cap C((0, T); [W^{2,p}(\Omega)]^3), \quad (3.23)$$

$$f^\epsilon, c^\epsilon \in C^1((0, T); W^{1,p}(\Omega)). \quad (3.24)$$

Proof. We will prove the existence of a local-in-time solution using the Banach contraction theorem. We first focus on the ODEs (3.1c) and (3.1d). We notice that relation (3.18) implies that $u_1^\epsilon, u_2^\epsilon, b^\epsilon \in W^{1,p}(\Omega)$, and since the functions $h_2 : \mathbb{R} \times \mathbb{R} \times \mathbb{R} \mapsto \mathbb{R}$, $h_3 : \mathbb{R} \times \mathbb{R} \mapsto \mathbb{R}$ and $h_4 : \mathbb{R} \mapsto \mathbb{R}$ are locally Lipschitz, we have by the property of superposition operator that the value of the functions h_2, h_3 and h_4 is also in $W^{1,p}(\Omega)$.⁵⁸ The space $W^{1,p}(\Omega)$ for $p > n$ is an algebra with pointwise multiplication, and thus it follows that the functions $(u_1^\epsilon, u_2^\epsilon, f^\epsilon, b^\epsilon) \mapsto P_1 = h_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon) f^\epsilon - \frac{\delta}{f_m} f^{\epsilon^2}$ and $(u_1^\epsilon, u_2^\epsilon, c^\epsilon, b^\epsilon) \mapsto P_2 = h_3(u_1^\epsilon, u_2^\epsilon) + c^\epsilon h_4(b^\epsilon)$ are

also $W^{1,p}$ -valued. Since the right-hand side functions of Eqs. (3.1c) and (3.1d) are locally Lipschitz, it follows from assumption (3.10) and embeddings (3.18)–(3.19) that the mapping $P : (W^{1,p}(\Omega))^5 \mapsto (W^{1,p}(\Omega))^2$, $P = (P_1, P_2)$, is a locally Lipschitz function.

For a fixed $T > 0$ we note that functions:

$$t \mapsto f_0 + \int_0^t P_1(u_1^\epsilon(s), u_2^\epsilon(s), f^\epsilon(s), b^\epsilon(s)) ds, \quad (3.25)$$

$$t \mapsto c_0 + \int_0^t P_2(u_1^\epsilon(s), u_2^\epsilon(s), c^\epsilon(s), b^\epsilon(s)) ds, \quad (3.26)$$

belong to the space $C([0, T]; W^{1,p}(\Omega))$.

Then, the system of the PDEs (3.1a), (3.1b) and (3.1e) with (3.3) can be rewritten as:

$$\begin{aligned} z_t &= Az + H(z), & \text{in } \Omega_T, \\ z(0, x) &= z_0(x), & \text{in } \Omega, \end{aligned} \quad (3.27)$$

with

$$z = \begin{pmatrix} u_1^\epsilon \\ u_2^\epsilon \\ b^\epsilon \end{pmatrix}, \quad A = \begin{pmatrix} A_1 & 0 & 0 \\ 0 & A_1 & 0 \\ 0 & 0 & A_3 \end{pmatrix},$$

and

$$H(z) = \begin{pmatrix} -\nabla u_1^\epsilon \cdot F_1[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] - u_1^\epsilon(\nabla \cdot F_1[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]) + G_1(z) \\ -\nabla u_2^\epsilon \cdot F_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] - u_2^\epsilon(\nabla \cdot F_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]) + h_1(z) \\ h_5(u_1^\epsilon, u_2^\epsilon) \end{pmatrix},$$

or, equivalently, we write that $A = A_1 \times A_2 \times A_3$ is sectorial in $X \times X \times X$, where $(A_1 \times A_2 \times A_3)(u_1^\epsilon, u_2^\epsilon, b^\epsilon) = (A_1 u_1^\epsilon, A_2 u_2^\epsilon, A_3 b^\epsilon)$,²⁷ and the mapping $H : (X^\gamma)^3 \mapsto (X)^3$, $H = (H_1, H_2, H_3)$ is defined as the mapping of the right-hand side of Eqs. (3.1a), (3.1b) and (3.1e). Using similar arguments as before for P , and assumption (3.9) we deduce that $H : (X^\gamma)^3 \mapsto (X)^3$ is locally Lipschitz continuous.

Let us now denote $Y = X \times X \times W^{1,p}(\Omega) \times W^{1,p}(\Omega) \times X$. For a fixed $T > 0$, we define the space

$$Y_T^\gamma = C([0, T]; Y^\gamma), \quad (3.28)$$

where $Y^\gamma = X^\gamma \times X^\gamma \times W^{1,p}(\Omega) \times W^{1,p}(\Omega) \times X^\gamma$ equipped with the norm $\|y\|_{Y^\gamma} = \max\{\|y_1\|_{X^\gamma}, \|y_2\|_{X^\gamma}, \|y_3\|_{W^{1,p}(\Omega)}, \|y_4\|_{W^{1,p}(\Omega)}, \|y_5\|_{X^\gamma}\}$, for $y = (y_1, y_2, y_3, y_4, y_5) \in Y^\gamma$.

We define the mapping $J : Y_T^\gamma \mapsto Y_T^\gamma$ with $J = (J_1, J_2, J_3, J_4, J_5)$ given by the following relation

$$J = \begin{cases} J_1 = e^{-A_1 t} u_{1_0} + \int_0^t e^{-A_1(t-s)} H_1(u_1^\epsilon(s), u_2^\epsilon(s), f^\epsilon(s), c^\epsilon(s), b^\epsilon(s)) ds, \\ J_2 = e^{-A_2 t} u_{2_0} + \int_0^t e^{-A_2(t-s)} H_2(u_1^\epsilon(s), u_2^\epsilon(s), f^\epsilon(s), c^\epsilon(s), b^\epsilon(s)) ds, \\ J_3 = f_0 + \int_0^t P_1(u_1^\epsilon(s), u_2^\epsilon(s), f^\epsilon(s), b^\epsilon(s)) ds, \\ J_4 = c_0 + \int_0^t P_2(u_1^\epsilon(s), u_2^\epsilon(s), c^\epsilon(s), b^\epsilon(s)) ds, \\ J_5 = e^{-A_3 t} b_0 + \int_0^t e^{-A_3(t-s)} H_3(u_1^\epsilon(s), u_2^\epsilon(s)) ds. \end{cases} \quad (3.29)$$

Let $R > 0$ be such that $\max\{\|u_{1_0}\|_{X^\gamma}, \|u_{2_0}\|_{X^\gamma}, \|b_0\|_{X^\gamma}\} < R/(2k_\gamma)$, where k_γ satisfies relation (3.22), and $\max\{\|f_0\|_{W^{1,p}(\Omega)}, \|c_0\|_{W^{1,p}(\Omega)}\} < R/2$ for $(u_{1_0}, u_{2_0}, f_0, c_0, b_0) \in Y^\gamma$. We define the ball

$$B_R = \{y \in Y_T^\gamma : \|y\|_{Y_T^\gamma} \leq R\} \subset Y_T^\gamma. \quad (3.30)$$

Thus, there exists $M_R > 0$ such that $\sup_{y \in B_R} \|(H_1, H_2, P_1, P_2, H_3)(y)\|_Y < M_R$. We show that J maps B_R into itself and that J is a strict contraction. By using relations (3.21)–(3.22) and (3.29) we obtain, for T small enough,

$$\begin{aligned} \|J_1[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon](t)\|_{X^\gamma} &\leq k_\gamma \|u_{1_0}\|_{X^\gamma} + M_R \int_0^T C_\gamma(t-s)^{-\gamma} e^{-\xi_1(t-s)} ds \\ &\leq \frac{R}{2} + M_R \frac{C_\gamma}{1-\gamma} T^{1-\gamma}. \end{aligned} \quad (3.31)$$

Similarly we obtain

$$\|J_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon](t)\|_{X^\gamma} \leq \frac{R}{2} + M_R \frac{C_\gamma}{1-\gamma} T^{1-\gamma} \quad (3.32)$$

and

$$\|J_5[u_1^\epsilon, u_2^\epsilon](t)\|_{X^\gamma} \leq \frac{R}{2} + M_R \frac{C_\gamma}{1-\gamma} T^{1-\gamma}. \quad (3.33)$$

Moreover, for J_3 and J_4 we have

$$\|J_l[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon](t)\|_{W^{1,p}(\Omega)} \leq \frac{R}{2} + M_R T, \quad l = 3, 4. \quad (3.34)$$

Hence, we can choose T sufficiently small such that $\{M_R \frac{C_\gamma}{1-\gamma} T^{1-\gamma}, M_R T\} < \frac{R}{2}$, to assert that $J(B_R) \subset B_R$. We note also that $J[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]$ is continuous from $[0, T]$ to E^γ , as it can be easily proved using inequality (3.22). Thus J maps B_R into itself.

If $y^1, y^2 \in B_R$ then for $t \in [0, T]$ we have

$$\begin{aligned} \|J_1[y^1](t) - J_1[y^2](t)\|_{X^\gamma} &\leq \int_0^t \|A_1^\gamma e^{-\xi_1(t-s)}\|_X \|H_1(y^1(s)) - H_1(y^2(s))\|_X ds \\ &\leq C_\gamma L_R \int_0^t (t-s)^{-\gamma} e^{\xi_1(t-s)} \|y^1 - y^2\|_{Y_T^\gamma}, \end{aligned}$$

where L_R is the Lipschitz constant of H . Similar estimates can be obtained for the differences of the rest of the arguments. Therefore, it follows that

$$\|J[y^1] - J[y^2]\|_{Y_T^\gamma} \leq \frac{1}{2} \|y^1 - y^2\|_{Y_T^\gamma} \quad \text{for all } y^1, y^2 \in B_R. \quad (3.35)$$

Hence for T small enough J is a contraction mapping. Therefore, by Banach fixed point theorem, J has a unique fixed point in B_R . Moreover, functions $f^\epsilon, c^\epsilon : [0, T] \mapsto C(\Omega)$ are locally Lipschitz, thus $f^\epsilon, c^\epsilon \in W^{1,\infty}([0, T]; W^{1,p}(\Omega))$ (see Theorem 4, Sec. 5.8.2 in Ref. 21). Therefore, it follows from Sec. 3.3 in Ref. 27 that there is a maximal time of existence T_{\max} of regular solution $(u_1^\epsilon, u_2^\epsilon, b^\epsilon) \in C([0, T_{\max}); (X^\gamma)^3)$ such that for $t \in (0, T_{\max})$ we have $(u_1^\epsilon, u_2^\epsilon, b^\epsilon) \in D(A)$. Then, from Ref. 27 (Sec. 3.3 and Theorem 3.5.2 in Sec. 3.5) we obtain that $(u_1^\epsilon, u_2^\epsilon, b^\epsilon) \in C^\zeta((0, T_{\max}); (X^\beta)^3)$ for some $\zeta, \beta \in (0, 1)$. Hence (3.1a), (3.1b) and (3.1e) are satisfied in a pointwise manner on $(0, T_{\max}) \times \Omega$. It follows now from relations (3.8) and (3.19), and the regularity theory of parabolic systems, that $u_1^\epsilon, u_2^\epsilon$ and b^ϵ are classical solutions of (3.1a), (3.1b) and (3.1e), respectively.

Let us now prove the uniqueness of solution. Let $y^1 = (u_1^1, u_2^1, f^1, c^1, b^1)$ and $y^2 = (u_1^2, u_2^2, f^2, c^2, b^2)$ be two solutions of system (3.1), with the same initial conditions. By linearity $y = y^1 - y^2$ is a solution of (3.1) with zero initial conditions. Then, since all nonlinear terms are Lipschitz continuous and the components of the solution are L^∞ -bounded functions on bounded time intervals, we have

$$\frac{d}{dt} \|y\|_{L^2(\Omega)}^2 \leq k_0 \|y\|_{L^2(\Omega)}^2, \quad \text{for } t \in [0, T_{\max}), \quad y(0) = 0, \quad (3.36)$$

for a constant k_0 , and since $\|y(0)\|_{L^2(\Omega)} = 0$, Gronwall's inequality implies that $\|y(t)\|_{L^2(\Omega)} = 0$ for all $t \geq 0$, so $y = 0$.

The equation for the ECM density given by (3.1c) does not involve any spatial derivatives and x behaves as a parameter. Thus, it is an ordinary differential equation in which the right-hand side is zero when $f^\epsilon(t, x) = 0$ and for which local Lipschitz conditions hold. Therefore, from Picard–Lindelöf theorem we obtain a local unique solution for the initial value problem (3.1c), with $f^\epsilon(0, x) = 0$. In the same way, we obtain a local unique solution for the initial value problem (3.1c), with $f^\epsilon(0, x) \geq 0$. Therefore, since $f^\epsilon(0, x) \geq 0$, from uniqueness of solutions we have $f^\epsilon(t, x) \geq 0$ for all $t > 0, x \in \Omega$. Then from maximum principle arguments it follows from system (3.27) that $u_1^\epsilon, u_2^\epsilon, b^\epsilon \geq 0$ on $[0, T_{\max}) \times \Omega$.

Finally, the equation for the integrin density given by (3.1d) can be treated in a similar manner as Eq. (3.1c) for the ECM density. Again we have an ordinary differential equation in which the right-hand side is greater than or equal to zero when

$c^\epsilon(t, x) = 0$, and for which local Lipschitz conditions hold. Therefore, $c^\epsilon(t, x) \geq 0$ on $[0, T_{\max}) \times \Omega$.

We proceed now with the proof of global-in-time solution. Let us first integrate Eq. (3.1a) on Ω . Then, from the boundary conditions (3.4)–(3.5) and relation (3.11) we obtain

$$\frac{d}{dt} \int_{\Omega} u_1^\epsilon(t, x) dx \leq B_1 |\Omega| - (M + D_1) \int_{\Omega} u_1^\epsilon(t, x) dx. \quad (3.37)$$

Thus, Gronwall's inequality yields the estimate

$$\sup_{t \in [0, T_{\max})} \|u_1^\epsilon(t, \cdot)\|_{L^1(\Omega)} \leq \max \left\{ \frac{B_1 |\Omega|}{M + D_1}, \|u_{10}\|_{L^1(\Omega)} \right\} := M_{u_1}. \quad (3.38)$$

Similarly, we have

$$\sup_{t \in [0, T_{\max})} \|u_2^\epsilon(t, \cdot)\|_{L^1(\Omega)} \leq \max \left\{ \frac{MM_{u_1} + B_2 |\Omega|}{D_2}, \|u_{20}\|_{L^1(\Omega)} \right\} := M_{u_2} \quad (3.39)$$

and

$$\sup_{t \in [0, T_{\max})} \|b^\epsilon(t, \cdot)\|_{L^1(\Omega)} \leq \max \left\{ \frac{\mu_1 M_{u_1} + \mu_2 M_{u_2}}{q_b}, \|b_0\|_{L^1(\Omega)} \right\} := M_b, \quad (3.40)$$

hence from relations (3.12)–(3.13) and Gronwall's inequality again we have

$$\sup_{t \in [0, T_{\max})} \|f^\epsilon(t, \cdot)\|_{L^1(\Omega)} \leq \max \left\{ \frac{\Lambda_1 M_b}{\Lambda_2}, \|f_0\|_{L^1(\Omega)} \right\} := M_f \quad (3.41)$$

and

$$\sup_{t \in [0, T_{\max})} \|c^\epsilon(t, \cdot)\|_{L^1(\Omega)} \leq \max \left\{ \frac{p_1 M_{u_1} + p_2 M_{u_2} + \Lambda_3 M_b}{\Lambda_4}, \|c_0\|_{L^1(\Omega)} \right\} := M_c. \quad (3.42)$$

Thus, from relations (3.6)–(3.9) we have for all $t \in [0, T_{\max})$:

$$\left\| \left(\sum_{j=1}^n \partial_{x_j} F_i[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] \right) + F_i[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] \right\|_{L^\infty(\Omega)} \leq L_N(\|u_1^\epsilon(t)\|_{L^1(\Omega)} + \|u_2^\epsilon(t)\|_{L^1(\Omega)} + \|f^\epsilon(t)\|_{L^1(\Omega)}) \leq L_N(M_{u_1} + M_{u_2} + M_f), \quad i = 1, 2. \quad (3.43)$$

From system (3.27), we rewrite the elliptic operators in the form:

$$\begin{aligned} & -D_u \Delta u_1^\epsilon + \sum_{j=1}^n b_1^j \partial_{x_j} u_1^\epsilon + d_1 u_1^\epsilon, \\ & -\epsilon \Delta u_2^\epsilon + \sum_{j=1}^n b_2^j \partial_{x_j} u_2^\epsilon + d_2 u_2^\epsilon, \\ & -D_b \Delta b^\epsilon + q_b b^\epsilon, \end{aligned}$$

where we denote by $b_i^j = F_{i,j}[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]$ and by $d_i = \sum_{j=1}^n \partial_{x_j} F_i[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon]$, $i = 1, 2, j = 1, \dots, n$. From relation (3.43) it follows that b_i^j and d_i , $i = 1, 2$, are

bounded on $(0, \infty) \times \Omega$. Hence, from the fact that the reaction terms are dissipative (see Ref. 13), it follows by Möser–Alikakos method (see Sec. 9.3 in Ref. 13) that the uniform in time $L^1(\Omega)$ estimate implies the uniform in time $L^\infty(\Omega)$ estimate for the solution $(u_1^\epsilon, u_2^\epsilon, b^\epsilon)$ of (3.27). Therefore, there is a constant M_∞ independent of ϵ (see Remark 3.1), such that

$$\sup_{t \in [0, T_{\max})} (\|u_1^\epsilon(t)\|_{L^\infty(\Omega)} + \|u_2^\epsilon(t)\|_{L^\infty(\Omega)} + \|b^\epsilon(t)\|_{L^\infty(\Omega)}) < M_\infty. \quad (3.44)$$

Moreover, from (3.12)–(3.13) and (3.44), and the comparison theorem it follows that

$$\sup_{t \in [0, T_{\max})} \|f^\epsilon(t)\|_{L^\infty(\Omega)} < \max \left\{ \|f_0\|_{L^\infty(\Omega)}, \frac{\Lambda_1 M_\infty}{\Lambda_2} \right\}, \quad (3.45)$$

and

$$\sup_{t \in [0, T_{\max})} \|c^\epsilon(t)\|_{L^\infty(\Omega)} < \max \left\{ \|c_0\|_{L^\infty(\Omega)}, (p_1 + p_2)M_\infty, \frac{\Lambda_3 M_\infty}{\Lambda_4} \right\}. \quad (3.46)$$

Relations (3.44)–(3.46) can be used to show that

$$\|H_i(u_1^\epsilon(s), u_2^\epsilon(s), f^\epsilon(s), c^\epsilon(s), b^\epsilon(s))\|_X \leq M_{\gamma_i} (1 + \|u_i^\epsilon(t)\|_{X^\gamma}) \quad \text{for } t \in [0, T_{\max}) \quad (3.47)$$

and

$$\|H_3(u_1^\epsilon(s), u_2^\epsilon(s))\|_X \leq M_{\gamma_3} \quad \text{for } t \in [0, T_{\max}), \quad (3.48)$$

where $M_{\gamma_i}, M_{\gamma_3}, i = 1, 2$, are constants depending on M_∞ .

We show now the global existence of solution by contradiction. Let us suppose that for $T_{\max} < \infty$ we have

$$\begin{aligned} \sup_{t \in [0, T_{\max})} (\|u_1^\epsilon(t)\|_{X^\gamma} + \|u_2^\epsilon(t)\|_{X^\gamma} + \|f^\epsilon(t)\|_{W^{1,p}(\Omega)} + \|c^\epsilon(t)\|_{W^{1,p}(\Omega)} \\ + \|b^\epsilon(t)\|_{X^\gamma}) \rightarrow \infty \quad \text{as } t \rightarrow T_{\max}. \end{aligned} \quad (3.49)$$

From relation (3.47) and Volterra-type integral inequality¹³ it follows, as in Corollary 3.3.5 in Ref. 27, that

$$\sup_{t \in [0, T_{\max})} \|u_i^\epsilon(t)\|_{X^\gamma} \leq \left(k_\gamma \|u_{i0}^\epsilon\|_{X^\gamma} + C_\gamma M_{\gamma_i} \int_0^{T_{\max}} \frac{e^{-\xi_i(t-s)}}{(t-s)^\gamma} \right) C_{T_{\max}},$$

where $C_{T_{\max}} := C_{C_\gamma M_{\gamma_i}, \gamma, T_{\max}}$, $i = 1, 2$, is a continuous function increasing with respect to T_{\max} , while for the function b^ϵ we have from relation (3.48) that

$$\sup_{t \in [0, T_{\max})} \|b^\epsilon(t)\|_{X^\gamma} \leq \left(k_\gamma \|b_0^\epsilon\|_{X^\gamma} + C_\gamma M_{\gamma_3} \int_0^{T_{\max}} \frac{e^{-\xi_3(t-s)}}{(t-s)^\gamma} \right).$$

Note that k_γ and C_γ are ϵ -independent constants since $0 < \epsilon \leq 1$ (see Theorem 1.3.4 in Ref. 27). We conclude that

$$\sup_{t \in [0, T_{\max})} (\|u_1^\epsilon(t)\|_{X^\gamma} + \|u_2^\epsilon(t)\|_{X^\gamma} + \|b^\epsilon(t)\|_{X^\gamma}) < \infty. \quad (3.50)$$

Whence, by (3.18) it follows now that

$$\sup_{t \in [0, T_{\max})} (\|u_1^\epsilon(t)\|_{W^{1,p}(\Omega)} + \|u_2^\epsilon(t)\|_{W^{1,p}(\Omega)} + \|b^\epsilon(t)\|_{W^{1,p}(\Omega)}) < \infty. \quad (3.51)$$

By Eq. (3.1c) and direct calculations, we obtain

$$\nabla f_t^\epsilon = (-\alpha \nabla u_1^\epsilon - \beta \nabla u_2^\epsilon + \theta_\beta \nabla b^\epsilon) f^\epsilon + h_6 \nabla f^\epsilon, \quad (3.52)$$

where

$$h_6 := h_2 - \frac{2\delta}{f_m} f^\epsilon = -\alpha u_1^\epsilon - \beta u_2^\epsilon + \theta_\beta b^\epsilon + \delta - \frac{2\delta}{f_m} f^\epsilon \leq \theta_\beta b^\epsilon + \delta. \quad (3.53)$$

For notational convenience, in what follows we denote various non-negative constants, which are independent of T or t , by c_j , $j = 1, 2, \dots, 8$.

Multiplying (3.52) by $p \nabla f^\epsilon |\nabla f^\epsilon|^{p-2}$, using (3.53), Young's inequality, and the estimates (3.44), (3.45) and (3.51), and integrating over Ω , we deduce that

$$\begin{aligned} \frac{d}{dt} \|\nabla f^\epsilon\|_{L^p(\Omega)}^p &\leq p \int_{\Omega} (-\alpha \nabla u_1^\epsilon - \beta \nabla u_2^\epsilon + \theta_\beta \nabla b^\epsilon) f^\epsilon \nabla f^\epsilon |\nabla f^\epsilon|^{p-2} dx \\ &\quad + p \theta_\beta \int_{\Omega} b^\epsilon |\nabla f^\epsilon|^p dx + p \delta \|\nabla f^\epsilon\|_{L^p(\Omega)}^p \\ &\leq p \max\{\alpha, \beta, \theta_\beta\} \|f^\epsilon\|_{L^\infty(\Omega)} \left(\int_{\Omega} |\nabla u_1^\epsilon| |\nabla f^\epsilon|^{p-1} dx \right. \\ &\quad \left. + \int_{\Omega} |\nabla u_2^\epsilon| |\nabla f^\epsilon|^{p-1} dx + \int_{\Omega} |\nabla b^\epsilon| |\nabla f^\epsilon|^{p-1} dx \right) \\ &\quad + p(\theta_\beta \|b^\epsilon\|_{L^\infty(\Omega)} + \delta) \|\nabla f^\epsilon\|_{L^p(\Omega)}^p \\ &\leq c_1 (\|\nabla u_1^\epsilon\|_{L^p(\Omega)}^p + \|\nabla u_2^\epsilon\|_{L^p(\Omega)}^p + \|\nabla b^\epsilon\|_{L^p(\Omega)}^p) + c_2 \|\nabla f^\epsilon\|_{L^p(\Omega)}^p. \end{aligned}$$

By Gronwall's inequality and previous estimates, we have

$$\sup_{t \in [0, T_{\max})} \|f^\epsilon(t)\|_{W^{1,p}(\Omega)} \leq c_3 e^{c_4 t}, \quad \text{for } t \geq 0. \quad (3.54)$$

Similarly, by Eq. (3.1d) and direct calculations, we obtain

$$\nabla c_t^\epsilon = p_1 \nabla u_1^\epsilon + p_2 \nabla u_2^\epsilon + p_3 \nabla b^\epsilon c^\epsilon + p_3 b^\epsilon \nabla c^\epsilon - q \nabla c^\epsilon. \quad (3.55)$$

Multiplying now (3.55) by $p \nabla c^\epsilon |\nabla c^\epsilon|^{p-2}$, using the estimates (3.44), (3.46) and (3.51), as well as Young's inequality, and integrating over Ω , we obtain

$$\begin{aligned} \frac{d}{dt} \|\nabla c^\epsilon\|_{L^p(\Omega)}^p &\leq p p_1 \int_{\Omega} \nabla u_1^\epsilon \nabla c^\epsilon |\nabla c^\epsilon|^{p-2} dx + p p_2 \int_{\Omega} \nabla u_2^\epsilon \nabla c^\epsilon |\nabla c^\epsilon|^{p-2} dx \\ &\quad + p p_3 \int_{\Omega} \nabla b^\epsilon c^\epsilon \nabla c^\epsilon |\nabla c^\epsilon|^{p-2} dx + p p_3 \int_{\Omega} b^\epsilon |\nabla c^\epsilon|^p dx - p q \int_{\Omega} |\nabla c^\epsilon|^p dx \\ &\leq p \max\{p_1, p_2, p_3\} \|c^\epsilon\|_{L^\infty(\Omega)} \left(\int_{\Omega} |\nabla u_1^\epsilon| |\nabla c^\epsilon|^{p-1} dx \right. \end{aligned}$$

$$\begin{aligned}
& + \int_{\Omega} |\nabla u_2^\epsilon| |\nabla c^\epsilon|^{p-1} dx + \int_{\Omega} |\nabla b^\epsilon| |\nabla c^\epsilon|^{p-1} dx \Big) \\
& + p(p_3 \|b^\epsilon\|_{L^\infty(\Omega)} - q) \|\nabla c^\epsilon\|_{L^p(\Omega)}^p \\
& \leq c_5 (\|\nabla u_1^\epsilon\|_{L^p(\Omega)}^p + \|\nabla u_2^\epsilon\|_{L^p(\Omega)}^p + \|\nabla b^\epsilon\|_{L^p(\Omega)}^p) + c_6 \|\nabla c^\epsilon\|_{L^p(\Omega)}^p.
\end{aligned}$$

This, together with Gronwall's inequality and previous estimates, yields

$$\sup_{t \in [0, T_{\max})} \|c^\epsilon(t)\|_{W^{1,p}(\Omega)} \leq c_7 e^{c_8 t}, \quad \text{for } t \geq 0. \quad (3.56)$$

Bounds (3.50), (3.54) and (3.56) contradict (3.49), therefore the solution exists globally. \square

Remark 3.1. It is easy to see how the proof of Möser–Alikakos method in Ref. 13 can be used in our case to show the ϵ -independent L^∞ -estimates given in relation (3.44). Following along the same lines with the proof of Lemma 9.3.1 in Ref. 13, we obtain a constant c' (as described in relation (9.3.11) of the proof in Ref. 13) such that

$$c' := \max\{\text{const. } a_0, 1, D|\Omega|, K^2\}, \quad (3.57)$$

where a_0 will be each of the diffusion coefficients D_u, ϵ and D_b . Using the fact that $0 < D_u, \epsilon, D_b \leq 1$ it follows that c' is ϵ -independent.

By this result, it follows that the rest of the estimates given by relations (3.45)–(3.48), (3.50)–(3.51), (3.54) and (3.56) are ϵ -independent.

3.2. Vanishing viscosity limit

Now we are ready to take the vanishing viscosity limit $\epsilon \rightarrow 0$ and prove the existence of solution for system (2.10). First we introduce the notion of a weak solution to problem (2.10)–(2.11) with (2.12)–(2.13).

Definition 3.1. A function $(u_1, u_2, f, c, b) \in L^\infty(\bar{\Omega}_T) \cap L^\infty(0, T; [L^1(\Omega)]^5)$ is called a weak solution of the problem (2.10)–(2.11) with (2.12)–(2.13) if:

(i) For all $\phi \in C_0^\infty(\Omega_T)$ we have

$$\begin{aligned}
& - \int_{\bar{\Omega}_T} u_1 \phi_t - D_u \nabla u_1 \cdot \nabla \phi + u_1 F_1[\underline{u}, f, c, b] \cdot \nabla \phi \\
& = \int_{\bar{\Omega}_T} [-Mu_1 + G_1(\underline{u}, b)] \phi,
\end{aligned} \quad (3.58)$$

$$- \int_{\bar{\Omega}_T} u_2 \phi_t + u_2 F_2[\underline{u}, f, c, b] \cdot \nabla \phi = \int_{\bar{\Omega}_T} [Mu_1 + G_2(\underline{u}, b)] \phi, \quad (3.59)$$

$$- \int_{\bar{\Omega}_T} f \phi_t = \int_{\bar{\Omega}_T} \left[-\alpha u_1 f - \beta u_2 f + \theta_\beta b f + \delta f \left(1 - \frac{f}{f_m} \right) \right] \phi, \quad (3.60)$$

$$- \int_{\bar{\Omega}_T} c \phi_t = \int_{\bar{\Omega}_T} [p_1 u_1 + p_2 u_2 + p_3 b c - q c] \phi, \quad (3.61)$$

$$- \int_{\bar{\Omega}_T} b \phi_t - D_b \nabla b \cdot \nabla \phi = \int_{\bar{\Omega}_T} [\mu_1 u_1 + \mu_2 u_2 - q_b b] \phi. \quad (3.62)$$

(ii) The functions u_1, u_2, f, c and b satisfy the initial conditions $u_{10}(x), u_{20}(x), f_0(x), c_0(x)$ and $b_0(x)$, respectively, given by (2.11), in the weak sense, i.e. there exists a set $E \subset [0, T]$ of Lebesgue measure zero such that $u_1(t_0, \cdot), u_2(t_0, \cdot), f(t_0, \cdot), c(t_0, \cdot)$ and $b(t_0, \cdot)$ are defined almost everywhere in Ω for $t_0 \in [0, T] \setminus E$ and satisfy:

$$\lim_{t_0 \rightarrow 0, t_0 \in [0, T] \setminus E} \int_{\Omega} |u_i(t_0, x) - u_{i0}(x)| dx = 0, \quad i = 1, 2, \quad (3.63)$$

$$\lim_{t_0 \rightarrow 0, t_0 \in [0, T] \setminus E} \int_{\Omega} |f(t_0, x) - f_0(x)| dx = 0, \quad (3.64)$$

$$\lim_{t_0 \rightarrow 0, t_0 \in [0, T] \setminus E} \int_{\Omega} |c(t_0, x) - c_0(x)| dx = 0, \quad (3.65)$$

$$\lim_{t_0 \rightarrow 0, t_0 \in [0, T] \setminus E} \int_{\Omega} |b(t_0, x) - b_0(x)| dx = 0. \quad (3.66)$$

We show now the L^1 -estimates with respect to time, which will be used in the proof of existence of solution to model (2.10).

Theorem 3.2. *Let the assumptions of Theorem 3.24 hold. Then for each $\rho > 0$ there exist nondecreasing functions $\omega_\rho^{u_i}, \omega_\rho^f, \omega_\rho^c, \omega_\rho^b \in C([0, \infty))$ with $\omega_\rho^{u_i}(0) = \omega_\rho^f(0) = \omega_\rho^c(0) = \omega_\rho^b(0) = 0$, $i = 1, 2$, such that for any $\epsilon \in (0, 1]$ and for any $t, t + \Delta t \in [0, T]$, $\Delta t \geq 0$ we have for a ball $B_\rho = \{|x| \leq \rho\}$ that:*

$$\int_{B_\rho} |u_i^\epsilon(t + \Delta t, x) - u_i^\epsilon(t, x)| dx \leq \omega_\rho^{u_i}(\Delta t), \quad i = 1, 2, \quad (3.67)$$

$$\int_{B_\rho} |f^\epsilon(t + \Delta t, x) - f^\epsilon(t, x)| dx \leq \omega_\rho^f(\Delta t), \quad (3.68)$$

$$\int_{B_\rho} |c^\epsilon(t + \Delta t, x) - c^\epsilon(t, x)| dx \leq \omega_\rho^c(\Delta t), \quad (3.69)$$

$$\int_{B_\rho} |b^\epsilon(t + \Delta t, x) - b^\epsilon(t, x)| dx \leq \omega_\rho^b(\Delta t). \quad (3.70)$$

Proof. Let us consider a function $g \in C_0^2(\Omega)$ with $\text{supp}(g) \subset B_\rho$. Then from the estimates obtained by Theorem 3.24 we have

$$\begin{aligned} & \left| \int_{\Omega} (u_2^\epsilon(t + \Delta t, x) - u_2^\epsilon(t, x))g(x)dx \right| \\ &= \left| \int_{\Omega} g(x) \int_t^{t+\Delta t} u_{2_t}^\epsilon(s, x)ds dx \right| \leq \left| \int_t^{t+\Delta t} \int_{\Omega} \epsilon u_2^\epsilon(s, x) \Delta g(x) dx ds \right| \\ &+ \left| \int_t^{t+\Delta t} \int_{\Omega} u_2^\epsilon(s, x) F_2[u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon] \nabla g(x) dx ds \right| \\ &+ \left| \int_t^{t+\Delta t} \int_{\Omega} [Mu_1^\epsilon(s, x) + G_2(u_1^\epsilon, u_2^\epsilon, b^\epsilon)]g(x) dx ds \right| \\ &\leq C(\rho, M_\infty) \Delta t \|g\|_{C^2(\Omega)}. \end{aligned}$$

Similarly we can obtain estimates for u_1, f, c and b . Then, as in Ref. 29, the L^1 -estimates of $u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon$ and b^ϵ with respect to time t follow. \square

Theorem 3.3. Let $u_i(0, x), b(0, x) \in X^\gamma, i = 1, 2$, and $f(0, x), c(0, x) \in W^{1,p}(\Omega)$. If assumptions of Theorem 3.24 are satisfied, then there exists a weak solution (u_1, u_2, f, c, b) of model (2.10) with (2.11)–(2.13), such that for all $T > 0$ the weak solution satisfies for almost all $(t, x) \in \bar{\Omega}_T$

$$0 \leq u_1(t, x), u_2(t, x), f(t, x), c(t, x), b(t, x) \leq C(M_\infty). \quad (3.71)$$

Proof. By Theorem 3.24, we have that for all $0 < \epsilon \leq 1$ and $T > 0$ there exists a classical solution $(u_1^\epsilon, u_2^\epsilon, f^\epsilon, c^\epsilon, b^\epsilon)$ of problem (3.1)–(3.3), which is uniformly bounded in $L^\infty(\bar{\Omega}_T)$. From estimates (3.51), (3.54) and (3.56) it follows that

$$\begin{aligned} & \sup_{t \in [0, T]} (\|u_1^\epsilon(t)\|_{W^{1,1}(\Omega)} + \|u_2^\epsilon(t)\|_{W^{1,1}(\Omega)} + \|f^\epsilon(t)\|_{W^{1,1}(\Omega)} \\ &+ \|c^\epsilon(t)\|_{W^{1,1}(\Omega)} + \|b^\epsilon(t)\|_{W^{1,1}(\Omega)}) < C. \end{aligned} \quad (3.72)$$

We consider for $m \in \mathbb{N}$ a sequence ϵ_m with $\epsilon_m \rightarrow 0$ for $m \rightarrow \infty$. Then by estimate (3.72) and L^1 -estimates with respect to time, obtained from Theorem 3.2, it follows by Fréchet–Kolmogorov theorem that the sequences $\{u_1^{\epsilon_m}, u_2^{\epsilon_m}, f^{\epsilon_m}, c^{\epsilon_m}, b^{\epsilon_m}\}$ are precompact in $L_{\text{loc}}^1(\bar{\Omega}_T)$. Using a standard diagonal extraction argument we obtain subsequences, denoted also as $\{u_1^{\epsilon_m}, u_2^{\epsilon_m}, f^{\epsilon_m}, c^{\epsilon_m}, b^{\epsilon_m}\}$, and functions $u_1, u_2, f, c, b \in L_{\text{loc}}^1(\bar{\Omega}_T)$ with $u_i^{\epsilon_m} \rightarrow u_i, i = 1, 2, f^{\epsilon_m} \rightarrow f, c^{\epsilon_m} \rightarrow c$ and $b^{\epsilon_m} \rightarrow b$ in $L_{\text{loc}}^1(\bar{\Omega}_T)$. This implies that the convergence is even pointwise a.e. for a suitable subsequence. From uniform L^1 -bounds of $\{u_1^{\epsilon_m}(t, \cdot), u_2^{\epsilon_m}(t, \cdot), f^{\epsilon_m}(t, \cdot), c^{\epsilon_m}(t, \cdot), b^{\epsilon_m}(t, \cdot)\}$ we have that $u_1(t, \cdot), u_2(t, \cdot), f(t, \cdot), c(t, \cdot), b(t, \cdot) \in L^1(\Omega)$. Multiplying now Eqs. (2.10a)–(2.10e) with a test function $\phi \in C_0^\infty(\Omega_T)$, taking the integral and integrating by

parts over $\bar{\Omega}_T$ yields

$$\begin{aligned} & - \int_{\bar{\Omega}_T} u_1^{\epsilon_m} \phi_t + D_u u_1^{\epsilon_m} \cdot \Delta \phi + u_1^{\epsilon_m} F_1[\underline{u}^{\epsilon_m}, f^{\epsilon_m}, c^{\epsilon_m}, b^{\epsilon_m}] \cdot \nabla \phi \\ & = \int_{\bar{\Omega}_T} [-M u_1^{\epsilon_m} + G_1(\underline{u}^{\epsilon_m}, b^{\epsilon_m})] \phi, \end{aligned} \quad (3.73)$$

$$\begin{aligned} & - \int_{\bar{\Omega}_T} u_2^{\epsilon_m} \phi_t + u_2^{\epsilon_m} F_2[\underline{u}^{\epsilon_m}, f^{\epsilon_m}, c^{\epsilon_m}, b^{\epsilon_m}] \cdot \nabla \phi \\ & = \int_{\bar{\Omega}_T} [M u_1^{\epsilon_m} + G_2(\underline{u}^{\epsilon_m}, b^{\epsilon_m})] \phi + \epsilon \int_{\bar{\Omega}_T} u_2^{\epsilon_m} \Delta \phi, \end{aligned} \quad (3.74)$$

$$\begin{aligned} & - \int_{\bar{\Omega}_T} f^{\epsilon_m} \phi_t = \int_{\bar{\Omega}_T} [-\alpha u_1^{\epsilon_m} f^{\epsilon_m} - \beta u_2^{\epsilon_m} f^{\epsilon_m} \\ & + \theta_\beta b^{\epsilon_m} f^{\epsilon_m} + \delta f^{\epsilon_m} (1 - f^{\epsilon_m}/f_m)] \phi, \end{aligned} \quad (3.75)$$

$$- \int_{\bar{\Omega}_T} c^{\epsilon_m} \phi_t = \int_{\bar{\Omega}_T} [p_1 u_1^{\epsilon_m} + p_2 u_2^{\epsilon_m} + p_3 b^{\epsilon_m} c^{\epsilon_m} - q c^{\epsilon_m}] \phi, \quad (3.76)$$

$$- \int_{\bar{\Omega}_T} b^{\epsilon_m} \phi_t + D_b b^{\epsilon_m} \cdot \Delta \phi = \int_{\bar{\Omega}_T} [\mu_1 u_1^{\epsilon_m} + \mu_2 u_2^{\epsilon_m} - q_b b^{\epsilon_m}] \phi. \quad (3.77)$$

The last term in (3.74) vanishes in the limit due to the uniform L^∞ -bound on $\{u_2^{\epsilon_m}\}$ and from the pointwise convergence the Lebesgue's dominated convergence theorem ensures that the limit (u_1, u_2, f, c, b) satisfies (3.58)–(3.62). Moreover, $(u_1, u_2, f, c, b) \in L^\infty(\bar{\Omega}_T)$, and by relations (3.44)–(3.46) and Remark 3.1 we obtain the bounds (3.71).

It remains to show the initial conditions (3.63)–(3.66). Let us first define the set $E \subset [0, T]$ such that for all $t_0 \in [0, T] \setminus E$ we have for almost all $x \in \Omega$ that (t_0, x) is Lebesgue point of u_1, u_2, f, c and b . The set E has Lebesgue measure zero. For any fixed $t_0 \in [0, T] \setminus E$ and $\rho > 0$ it follows from Theorem 3.2 that

$$\int_{B_\rho} |u_i(t_0, x) - u_{i_0}(x)| dx \leq \int_{B_\rho} |u_i(t_0, x) - u_i^{\epsilon_m}(t_0, x)| dx + \omega_\rho^{u_i}(t_0), \quad i = 1, 2.$$

The pointwise convergence of $\{u_i^{\epsilon_m}\}$, $i = 1, 2$, yields

$$\int_{B_\rho} |u_i(t_0, x) - u_{i_0}(x)| dx \leq \omega_\rho^{u_i}(t_0), \quad i = 1, 2.$$

The properties of $\omega_\rho^{u_i}(t_0)$, $i = 1, 2$, give (3.63) since u_i has compact support. Similarly we obtain relations (3.64)–(3.66). \square

4. Numerical Results

4.1. Non-dimensionalisation of the model

In this section, we investigate numerically the type of patterns exhibited by model (2.10) in the one-dimensional case. Let $R_s > 0$ be the cells sensing radius (i.e. the

maximum range over which cells can detect other surrounding cells). We consider a bounded domain $\Omega = [0, R_s]$, and following the approach in Ref. 24, we choose the non-local terms $F_i[\underline{u}, f, c, b]$, $i = 1, 2$, to be given by

$$F_i[\underline{u}, f, c, b](t, x) := \frac{1}{R_s} \int_0^{R_s} \sum_{k=0}^1 \eta(k) K(r) g_i(\underline{v}(t, x + r\eta(k)), c(t, x), b(t, x)) dr, \quad (4.1)$$

where $\eta(k) = (-1)^k$, $k = 0, 1$ and g_i , $i = 1, 2$, as described in Sec. 2 (see relation (2.5)).

Let us define the kernel K , assuming that it is attractive at medium/long ranges (i.e. at the edges of the cell) and repulsive at very short ranges (i.e. over cell surface), and thus can be defined as

$$K(x) := q_a K_a(x) - q_r K_r(x), \quad (4.2)$$

with q_a and q_r describing the magnitudes of attractive and repulsive interactions, respectively, and $K_a(x)$ and $K_r(x)$ describe the spatial ranges over which these interactions take place. We consider translated Gaussian attraction and repulsion kernels (as in Ref. 20):

$$K(x) = \frac{q_a}{\sqrt{2\pi m_a^2}} e^{-\frac{(x-s_a)^2}{2m_a^2}} - \frac{q_r}{\sqrt{2\pi m_r^2}} e^{-\frac{(x-s_r)^2}{2m_r^2}}, \quad (4.3)$$

where s_a and s_r represent half of the length of attraction and repulsion ranges, respectively, with $s_r < s_a$. Also, $m_j = s_j/8$, $j = a, r$, represent the width of the attractive and the repulsive interaction ranges.

To perform numerical simulations, we first non-dimensionalise system (2.10) by using the following quantities:

$$\begin{aligned} \tilde{t} &= \frac{t}{\tau}, \quad \tilde{x} = \frac{x}{L_0}, \quad \tilde{u}_i = \frac{u_i}{k_u}, \quad \tilde{f} = \frac{f}{f_m}, \\ \tilde{c} &= \frac{c}{c_m}, \quad \tilde{b} = \frac{b}{b_m}, \quad \tilde{R}_s = \frac{R_s}{L_0}, \quad \tilde{r} = \frac{r}{L_0}, \\ \tilde{S}(\tilde{c}, \tilde{b}) &= \frac{\tau k_u}{L_0^2} S(c_m \tilde{c} + b_m \tilde{b}), \quad \tilde{S}_i(\tilde{c}, \tilde{b}) = \frac{\tau k_u}{L_0^2} S_i(c_m \tilde{c} + b_m \tilde{b}), \\ \tilde{C}_i(\tilde{c}, \tilde{b}) &= \frac{\tau f_m}{L_0^2} C_i(c_m \tilde{c} + b_m \tilde{b}), \quad i = 1, 2. \end{aligned} \quad (4.4)$$

The length scale, L_0 , is in the range of 0.1–1 cm, and is defined as the maximum invasion distance of the cancer cells at the early stage of invasion.² The time scale is defined as $\tau := L_0^2/D_\tau$, where D_τ is the characteristic diffusion coefficient ($\sim 10^{-6} \text{ cm}^2 \text{ s}^{-1}$). Furthermore, we rescale the cancer cells, the ECM, the integrins and the TGF- β with k_u , f_m , c_m and b_m , respectively. Here, k_u is the carrying capacity of the cancer cell populations and it is taken to be $\sim 6.7 \cdot 10^7 \text{ cell/volume}$, and f_m is the maximum ECM density at which the ECM fills up all available physical space and it is taken to be equal to 4 mg/volume, as in Ref. 19. Finally, c_m

is the maximum integrin density and it is taken to be $5 \cdot 10^4$ integrins per cell (as in Ref. 7), while b_m is the maximum TGF- β concentration taken to be equal to 141.59 ng/volume (as in Ref. 34).

We choose the dimensionless functions $\tilde{K}(\tilde{r}) := L_0 K(L_0 \tilde{r}) = L_0 K(r)$ and $\tilde{g}_i(\underline{\tilde{u}}, \tilde{f}, \tilde{c}, \tilde{b}) := (\tau/L_0^2) g_i(\underline{u}, f, c, b)$, $i = 1, 2$. Therefore, the non-local terms are given by $\tilde{F}_i[\underline{\tilde{u}}, \tilde{f}, \tilde{c}, \tilde{b}] := (\tau/L_0) F_i[\underline{u}, f, c, b]$, $i = 1, 2$.

Finally, we obtain the dimensionless parameters:

$$\begin{aligned} \tilde{D}_u &= \frac{D_u}{D_\tau}, \quad \tilde{D}_b = \frac{D_b}{D_\tau}, \quad \tilde{M} = \tau M, \quad \tilde{\alpha} = \tau \alpha k_u, \quad \tilde{\beta} = \tau \beta k_u, \\ \tilde{\theta}_\beta &= \tau \theta_\beta b_m, \quad \tilde{\delta} = \tau \delta, \quad \tilde{p}_3 = \tau p_3 b_m, \quad \tilde{q} = \tau q, \quad \tilde{q}_b = \tau q_b, \\ \tilde{r}_i &= \tau r_i, \quad \tilde{p}_i = \frac{\tau p_i k_u}{c_m} \quad \text{and} \quad \tilde{\mu}_i = \frac{\tau \mu_i k_u}{b_m}, \quad i = 1, 2. \end{aligned} \quad (4.5)$$

After dropping the tildes for notational convenience, we obtain the following non-dimensionalised system:

$$\begin{aligned} \frac{\partial u_1}{\partial t} &= D_u \frac{\partial^2 u_1}{\partial x^2} - \frac{\partial}{\partial x} (u_1 F_1[\underline{u}, f, c, b]) \\ &\quad - M u_1 + r_1 u_1 (1 - u_1 - u_2) (1 - c_b b), \end{aligned} \quad (4.6a)$$

$$\frac{\partial u_2}{\partial t} = - \frac{\partial}{\partial x} (u_2 F_2[\underline{u}, f, c, b]) + M u_1 + r_2 u_2 (1 - u_1 - u_2) (1 + c_b b), \quad (4.6b)$$

$$\frac{\partial f}{\partial t} = -\alpha u_1 f - \beta u_2 f + \theta_\beta b f + \delta f (1 - f), \quad (4.6c)$$

$$\frac{\partial c}{\partial t} = p_1 u_1 + p_2 u_2 + p_3 b c - q c, \quad (4.6d)$$

$$\frac{\partial b}{\partial t} = D_b \frac{\partial^2 b}{\partial x^2} + \mu_1 u_1 + \mu_2 u_2 - q_b b. \quad (4.6e)$$

4.2. Pattern formation

To discretize our model we use a time-splitting approach. We use a Crank–Nicolson scheme to propagate the solution of the diffusion term. Then, we use the Nessyahu–Tadmor scheme⁵³ for the time-propagation of the advection terms. Finally, for the time-propagation of the reaction terms we use a fourth-order Runge–Kutta algorithm, where the integrals are further discretised using the Simpson’s rule. All simulations are performed on a domain of length $L = 10$ with periodic boundary conditions (introduced to approximate the dynamics on an infinite domain). For this reason, the integrals are wrapped-up at the boundaries. The simulations ran for times up to $t = 1000$, but for clarity in Figs. 2–5 we show mainly the dynamics for $t \leq 400$. If the patterns do not reach a steady state before $t = 400$, we add inset figures showing the dynamics for $t \leq 1000$.

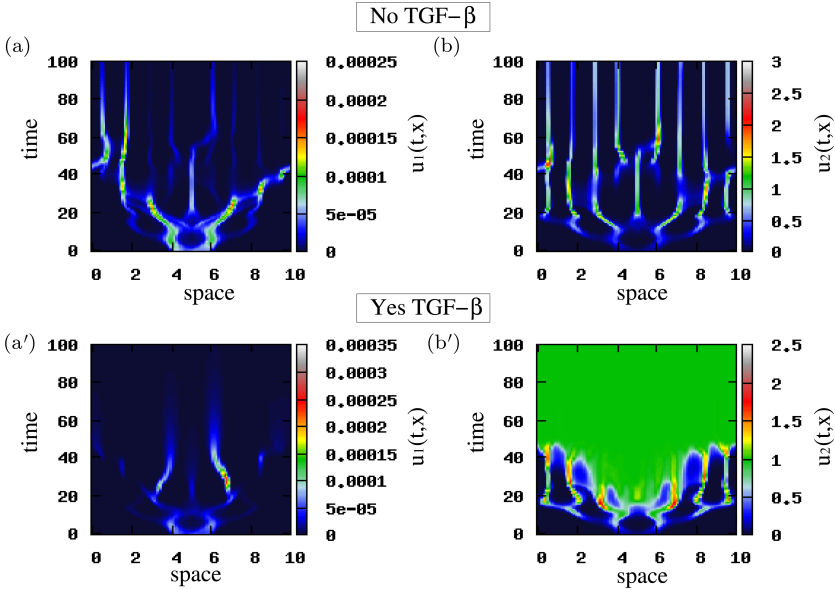


Fig. 2. (Colour online) Patterns exhibited by model (4.6) showing the effect of TGF- β on cancer cell density for cell-matrix adhesion greater than cell-cell adhesion, i.e. $s_1^* = 1.8, s_2^* = 0.6, s^* = 1, c_1^* = 1.9$ and $c_2^* = 2.5$. The rest of model parameters are given in Table A.2. (a), (b) Density of u_1 and u_2 populations in the absence of TGF- β ; (a'), (b') Density of u_1 and u_2 populations in the presence of TGF- β .

The initial conditions for the cancer cell populations are small random perturbations of rectangular-shaped aggregations located in the middle of the domain

$$u_i(0, x) = \begin{cases} u_i^c + \text{rand}(0, 10^{-4}), & x \in (L/2 - 1, L/2 + 1), \\ 0, & \text{everywhere else,} \end{cases} \quad (4.7)$$

with $u_1^c = 0$ and $u_2^c = 0.1$. For the ECM density, f , we assume that the tumour has already degraded some of its surrounding tissues:

$$f(0, x) = 1 - 0.5u_1(0, x) - 0.5u_2(0, x). \quad (4.8)$$

Finally, the integrin density and TGF- β concentration, c and b , respectively, are proportional to the initial tumour cell density

$$c(0, x) = 0.5u_1(0, x) + 0.5u_2(0, x) \quad (4.9)$$

and

$$b(0, x) = 0.05u_1(0, x) + 0.05u_2(0, x). \quad (4.10)$$

To investigate the effect of TGF- β signalling on cell proliferation, movement and aggregation (the last two aspects being controlled by cell adhesion), we focus on three possible cases for the magnitudes of cell-cell and cell-matrix adhesion. For each of these three cases, we investigate the dynamics of u_1 and u_2 populations

when TGF- β is absent and does not influence cell proliferation or cell adhesion (i.e. for $c_b = a_{b_i} = d_b = e_{b_i} = \theta_\beta = p_3 = \mu_1 = \mu_2 = 0, i = 1, 2$), and when TGF- β is present and influences both cell proliferation and cell adhesion (i.e. for $c_b = 20$ and $a_{b_i}, d_b, e_{b_i}, \theta_\beta, p_3, \mu_1, \mu_2, \neq 0, i = 1, 2$).

- (i) *Cell-cell adhesion < cell-matrix adhesion.* To investigate the effect of greater cell-matrix adhesion, we choose $s_1^* = 1.8, s_2^* = 0.6, s^* = 1, c_1^* = 1.9$ and $c_2^* = 2.5$ and the rest of model parameters as given in Table A.2. We see in Figs. 2(a) and 2(b) that in the absence of TGF- β , the population of early stage cancer cells (u_1) decreases, while the population of late stage cancer cells (u_2) increases and dominates the long-term dynamics. This behaviour is expected due to the mutation term “ $-Mu_1$ ”, and due to large cell-matrix adhesion, which impedes cells to move and thus leads to the formation of stationary pulses for $t > 50$. Considering now the effect of TGF- β , we see in Figs. 2(a') and 2(b') that population u_1 vanishes faster, due to the presence of antiproliferative and proapoptotic signals from TGF- β (described by $c_b > 0$ in Eq. (4.6a)). Population u_2 persists and increases significantly, due to the promoting effects of TGF- β on the late stages of cancer, which also induces the movement of the cancer cells (via EMT) thus leading to their spread over the domain until they reach the boundaries.

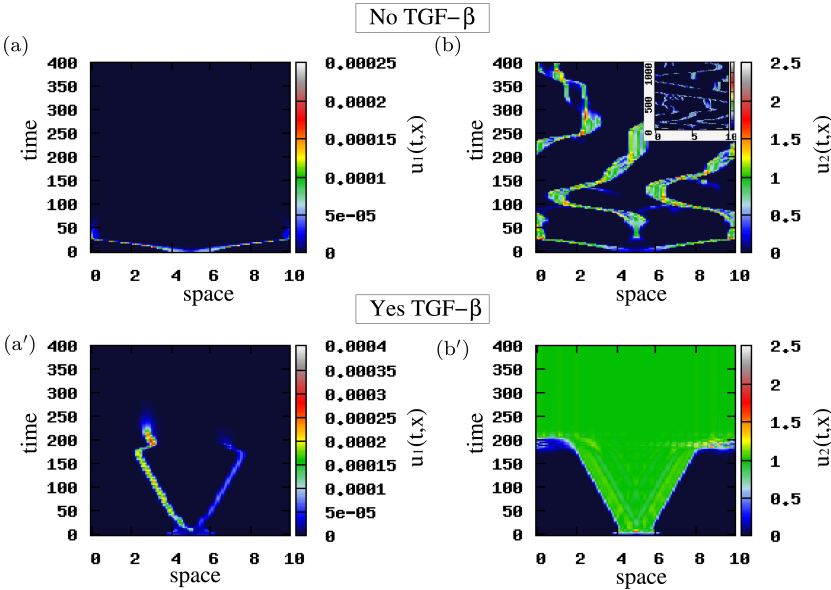


Fig. 3. (Colour online) Patterns exhibited by model (4.6) showing the effect of TGF- β on cancer cell density for cell-cell adhesion greater than cell-matrix adhesion, i.e. $s_1^* = 2.4, s^* = 2.1, s_2^* = 2, c_1^* = 1.1$ and $c_2^* = 0.9$. The rest of model parameters are given in Table A.2. (a), (b) Density of u_1 and u_2 populations in the absence of TGF- β . The inset in panel (b) shows the long-term dynamics of $u_2(t, x)$ (for $t \leq 1000$); (a'), (b') Density of u_1 and u_2 populations in the presence of TGF- β .

- (ii) *Cell-cell adhesion* > *cell-matrix adhesion*. To investigate the effect of greater cell-cell adhesion, we choose $s_1^* = 2.4$, $s^* = 2.1$, $s_2^* = 2$, $c_1^* = 1.1$ and $c_2^* = 0.9$, and the rest of model parameters as given in Table A.2. We see in Figs. 3(a) and 3(b) that due to the weak cell-matrix adhesive forces, u_1 and u_2 cells start to move through the domain in a collective manner. Figures 3(a) and 3(a') show that u_1 population vanishes in the absence and in the presence of TGF- β (due to the mutation term). We also note that the spread of u_1 cells is reduced in the presence of TGF- β , likely due to the positive effect of TGF- β on cell-matrix adhesion (see the term “ $+e_{b_i}b$ ” in Eqs. (2.6)). In Figs. 3(b) and 3(b') we see that the u_2 population changes its movement from a chaotic-like dynamics (in the absence of TGF- β ; panel (b)) to a spread over the whole domain (in the presence of TGF- β ; panel (b')), as a result of a decrease in the cell-cell adhesion induced by the tumour growth factor.
- (iii) *Cell-cell adhesion* = *cell-matrix adhesion*. To ensure the same values for the adhesive strength functions (2.6) when there is no TGF- β in the system, we choose $s_i^* = s^* = c_i^* = 0.8$ and $a_i = d = e_i = 0.5$, $i = 1, 2$. In Figs. 4(a) and 4(b), we see that some cells in the two cancer sub-populations move quickly to the left and the right, reaching the boundaries, while other cells (both u_1 and u_2) stay in the middle of the domain and create a chaotic-like pattern (even if u_1 is slowly eliminated for $t > 50$). If we now add TGF- β to the system, we see

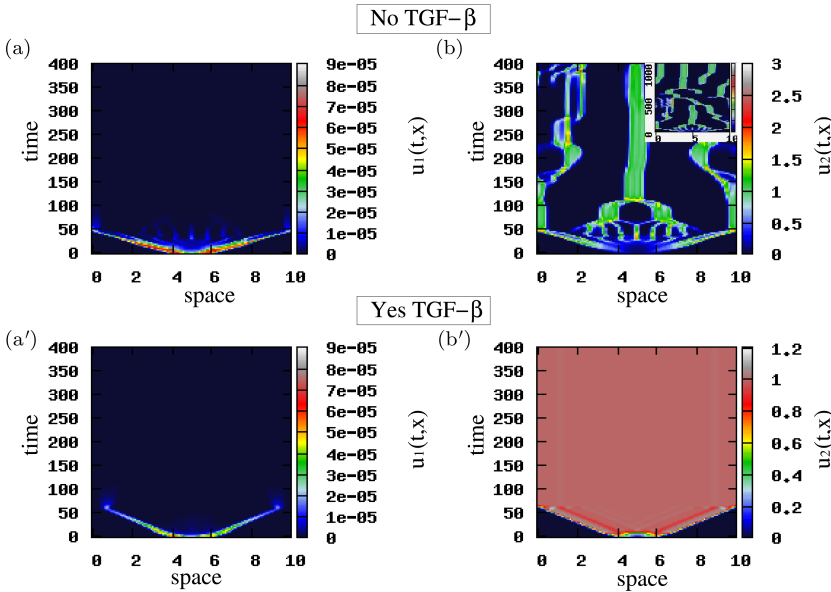


Fig. 4. (Colour online) Patterns exhibited by model (4.6) showing the effect of TGF- β on cancer cell density for the same cell-cell and cell-matrix adhesion, i.e. $s_i^* = s^* = c_i^* = 0.8$ and $a_i = d = e_i = 0.5$, $i = 1, 2$. The rest of model parameters are given in Table A.2. (a), (b) Density of u_1 and u_2 populations in the absence of TGF- β . The inset in panel (b) shows the long-term dynamics of $u_2(t, x)$ (for $t \leq 1000$); (a'), (b') Density of u_1 and u_2 populations in the presence of TGF- β .

in Figs. 4(a') and 4(b') that both u_1 and u_2 populations move slower towards the edges of the domain. In contrast to the u_2 population in the absence of TGF- β , which exhibits a chaotic clumping and splitting behaviour (panel (b)), the u_2 population in the presence of TGF- β exhibits travelling-wave dynamics (panel (b')). This is different from the dynamics observed in Fig. 3(b') where the u_2 cells move in a travelling-wave manner up to $t = 200$, after which they quickly move towards the boundaries.

Reducing now the magnitudes of cellular adhesion forces to $s_i^* = s^* = c_i^* = 0.1$, $i = 1, 2$, we see in Fig. 5 that irrespective of the absence/presence of TGF- β , population u_1 forms a stationary aggregation that eventually vanishes for large times, while population u_2 exhibits a travelling wave. This behaviour might be explained by the combined effect of high mutation rate and clonal competition (see the logistic growth terms in (2.2)), since adhesive forces are very small and lead to the spread of population u_2 . In contrast to the dynamics in Figs. 3(b) and 3(b'), and 4(b) and 4(b'), where the u_2 cells seem to travel slower towards the boundaries in the presence of TGF- β (compared to the absence of TGF- β), in Fig. 5 the u_2 cells travel faster to the boundaries in the presence of TGF- β . We deduce from here that the spread of tumour cells depends both on the magnitude of adhesive forces as well as on the presence of TGF- β molecules.

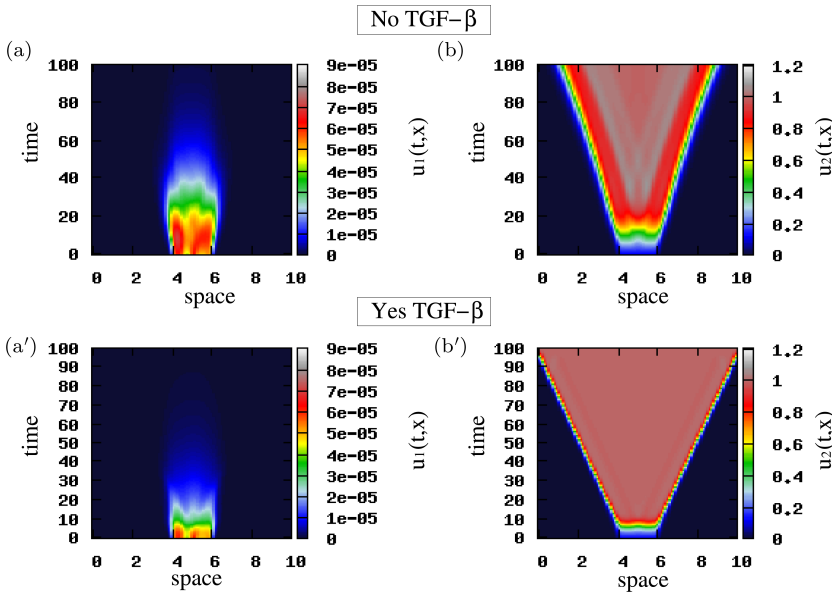


Fig. 5. (Colour online) Patterns exhibited by model (4.6) showing the effect of TGF- β on cancer cell density for the same cell-cell and cell-matrix adhesion, i.e. $s_i^* = s^* = c_i^* = 0.1$ and $a_i = d = e_i = 0.5$, $i = 1, 2$. The rest of model parameters are given in Table A.2. (a), (b) Density of u_1 and u_2 populations in the absence of TGF- β ; (a'), (b') Density of u_1 and u_2 populations in the presence of TGF- β .

We note here that we also investigated numerically the case when TGF- β is present, but does not influence cell proliferation or cell adhesion (i.e. for $\theta_\beta, p_3, \mu_1, \mu_2 \neq 0$, and $c_b = a_{b_i} = d_b = e_{b_i} = 0$, $i = 1, 2$). The patterns (not shown here) that we obtained were similar to those presented in Figs. 2(a) and 2(b), 3(a) and 3(b), 4(a) and 4(b), 5(a) and 5(b). This suggests that the effect of TGF- β on the cancer cell density is greater than the effect on the ECM and on the integrins density.

5. Conclusion and Discussion

In this paper, we introduced a model of integro-differential equations describing the dynamics of early stage and late stage cancer cell populations, under the effect of TGF- β signalling. The model was then used to investigate the role of TGF- β on cellular adhesion and proliferation.

We first proved the global existence of bounded solutions to our non-local model by taking a vanishing viscosity approach and approximating our model with a non-local parabolic PDE. The proof used the Banach contraction mapping theorem, the Möser–Alikakos method and the vanishing viscosity method.

We then investigated numerically the solution of this non-local model, paying particular attention to the effect of TGF- β on cell–cell and cell–matrix interactions. We showed that: (i) In the absence of TGF- β , the magnitudes of cell–cell and cell–matrix interactions influenced the formation of cancer cell aggregation at specific position in space (see Figs. 2(b), 3(b) and 4(b)); (ii) The consideration of TGF- β leads to the spread of mutated (i.e. u_2) cancer cells over the whole domain mainly in a travelling-wave manner (with no cell aggregations; see Figs. 3(b'), 4(b') and 5(b')). We also emphasise that the speed at which cells spread depended on the presence/absence of TGF- β and on the magnitudes of cell adhesion forces (see Figs. 4(b) and 4(b') versus Figs. 5(b) and 5(b')).

While the numerical investigation of cancer spread uncovered an interesting combined effect of cell adhesion and presence/absence of TGF- β , in the future we plan to investigate analytically the travelling waves and study the effect of parameters related to TGF- β and cell adhesion on the speed of these waves.

Appendix A. Summary of Model Variables and Parameters

Here we present two tables with the model variables and parameters. In Table A.1, we list the model variables with their units. In Table A.2, we list the parameters of our model and their corresponding units and non-dimensional values used in the simulations.

Parameter estimation

- Attraction and repulsion ranges were chosen to be smaller or equal to sensing radius, with the repulsion range to be smaller than the attraction range.²⁵

Table A.1. A list of model variables with their units. Since we are in 1D, length and volume coincide and we express the variables in terms of domain length.

Variable	Description	Dimensional units
u_1	Early stage cancer cell density	cell/length
u_2	Late stage cancer cell density	cell/length
f	ECM density	mg/length
c	Integrin density	integrins/cell
b	TGF- β concentration	mg/length

Table A.2. A list of model parameters with their units and their non-dimensional values, obtained from (4.4) and (4.5), which we used during numerical simulations.

Param.	Description	Dimensional units	Non-dim. value (\bar{p})	Reference
D_u	Diffusion coefficient of u_1	length ² /time	0.0001	11
R_s	Sensing radius	length	0.99	3 and 24
q_a	Magnitude of attraction	length ² /cell	0.09	Estimated
q_r	Magnitude of repulsion	length ² /cell	0.01	Estimated
s_a	Attraction range	length	0.99	Estimated
s_r	Repulsion range	length	0.25	Estimated
m_a	Width of attraction kernel	length	0.99/8	Estimated
m_r	Width of repulsion kernel	length	0.25/8	Estimated
r_1	Growth rate of u_1	1/time	0.1	50
r_2	Growth rate of u_2	1/time	0.2	50
M	Mutation rate	1/time	0.05	14, 28 and 43
c_b	Coeff. related to the effect of TGF- β on cancer cell proliferation	Nondim.	20	Estimated
a_1	Coeff. related to the number of integrins necessary for max self-adhesion between u_1	cell/integrins	0.7	Estimated
a_2	Coeff. related to the number of integrins necessary for max self-adhesion between u_2	cell/integrins	0.3	Estimated
d	Coeff. related to the number of integrins necessary for max cell-cell cross-adhesion	cell/integrins	0.5	Estimated
e_1	Coeff. related to the number of integrins necessary for max cell-ECM adhesion for u_1	cell/integrins	1.8	Estimated
e_2	Coeff. related to the number of integrins necessary for max cell-ECM adhesion for u_2	cell/integrins	2.5	Estimated

(Continued)

Table A.2. (Continued)

Param.	Description	Dimensional units	Non-dim. value (\tilde{p})	Reference
a_{b_1}	Coeff. related to the effect of TGF- β on self-adhesion between u_1 cells	length/mg	0.5	Estimated
a_{b_2}	Coeff. related to the effect of TGF- β on self-adhesion between u_2 cells	length/mg	0.3	Estimated
d_b	Coeff. related to the effect of TGF- β on cell-cell cross-adhesion	length/mg	0.4	Estimated
e_{b_1}	Coeff. related to the effect of TGF- β on cell-ECM adhesion for u_1 cells	length/mg	0.8	Estimated
e_{b_2}	Coeff. related to the effect of TGF- β on cell-ECM adhesion for u_2 cells	length/mg	0.9	Estimated
s_1^*	Magnitude of self-adhesion forces of u_1	length/(time \cdot cell)	0.1–2.4	Estimated
s_2^*	Magnitude of self-adhesion forces of u_2	length/(time \cdot cell)	0.1–2	Estimated
s^*	Magnitude of cross-adhesion forces	length/(time \cdot cell)	0.1–2.1	Estimated
c_1^*	Magnitude of cell-ECM forces of u_1	length/(time \cdot cell)	0.1–1.9	Estimated
c_2^*	Magnitude of cell-ECM forces of u_2	length/(time \cdot cell)	0.1–2.5	Estimated
α	Rate of ECM degradation by u_1	length/(time \cdot cell)	1	59
β	Rate of ECM degradation by u_2	length/(time \cdot cell)	2	59
θ_β	Binding rate of TGF- β to ECM components	length/(time \cdot mg)	0.77	Estimated
δ	ECM remodelling rate	1/time	0.25	11
p_1	Production rate of c by u_1	integrins/(time \cdot cell)	0.05	Estimated
p_2	Production rate of c by u_2	integrins/(time \cdot cell)	0.1	Estimated
p_3	Up-regulation rate of c by b	length/(time \cdot mg)	0.2	Estimated
q	Decay rate of c	1/time	0.3	39
D_b	Diffusion coefficient of TGF- β	length ² /time	0.007	64
μ_1	Production rate of b by u_1	mg/(time \cdot cell)	0.05	36 and 69
μ_2	Production rate of b by u_2	mg/(time \cdot cell)	0.05	36 and 69
q_b	Decay rate of b	1/time	0.05	39

- Various experimental studies^{15,50} have shown that doubling times for tumour cells range from 1–10 days. This corresponds to growth rates between $(\ln(2)/10, \ln(2)/1) = (0.07, 0.7)$. In this study, we assume that $\tilde{r}_1 = 0.1$ and $\tilde{r}_2 = 0.2$.
- Experimental studies^{14,28,43} have shown that the mutation rate ranges between $M = 10^{-3}/\text{day}$ and $M = 0.1/\text{day}$. Thus the non-dimensional value of the mutation rate is in the range between $\tilde{M} = 0.001$ and $\tilde{M} = 0.1$ (for highly aggressive tumours). In this study, we choose $\tilde{M} = 0.05$.
- The parameters $a_i, d, e_i, s_i^*, s^*, c_i^*, i = 1, 2$, were based on the range of the adhesion strength parameters used in Ref. 3.
- Experimental studies³⁵ have shown greater production of integrins for mutated cancer cells. Thus, we choose $p_1 < p_2$.
- Experimental studies^{16,17,39,40} have shown that the half-lives of the integrins range from 0.04–4 days. This corresponds to a decay rate between $(\ln(2)/4, \ln(2)/0.04) = (0.17, 17.3)$. In this study, we assume that $\tilde{q} = 0.3$.
- The remodelling rate was chosen to be greater than cell proliferation rate, as considered also in Ref. 11.

Acknowledgments

The authors would like to thank Dr. D. Trucu for fruitful discussions. V.B. acknowledges support from an Engineering and Physical Sciences Research Council (UK) Grant Number EP/L504932/1. R.E. was partially supported by an Engineering and Physical Sciences Research Council (UK) Grant Number EP/K033689/1.

References

1. D. Ambrosi and L. Preziosi, On the closure of mass balance models for tumor growth, *Math. Models Methods Appl. Sci.* **12** (2002) 737–754.
2. A. R. A. Anderson, M. A. J. Chaplain, E. L. Newman, R. J. C. Steele and A. M. Thompson, Mathematical modelling of tumour invasion and metastasis, *J. Theor. Med.* **2** (2000) 129–154.
3. N. J. Armstrong, K. J. Painter and J. A. Sherratt, A continuum approach to modelling cell–cell adhesion, *J. Theor. Biol.* **243** (2006) 98–113.
4. G. Ascolani and P. Liò, Modeling TGF- β in early stages of cancer tissue dynamics, *PLoS One* **9** (2014) e88533.
5. N. Bellomo, A. Bellouquid and N. Chouhad, From a multiscale derivation of nonlinear cross-diffusion models to Keller–Segel models in a Navier–Stokes fluid, *Math. Models Methods Appl. Sci.* **26** (2016) 2041–2069.
6. N. Bellomo, N. Li and P. K. Maini, On the foundations of cancer modelling: Selected topics, speculations and perspectives, *Math. Models Methods Appl. Sci.* **18** (2008) 593–646.
7. S. Benedetto, R. Pulito, S. G. Crich, G. Tarone, S. Aime, L. Silengo and J. Hamm, Quantification of the expression level of integrin receptor $\alpha v \beta 3$ in cell lines and MR imaging with antibody-coated iron oxide particles, *Magn. Reson. Med.* **56** (2006) 711–716.

8. H. Byrne and L. Preziosi, Modelling solid tumour growth using the theory of mixtures, *Math. Med. Biol.* **20** (2003) 341–366.
9. F. Calvo and E. Sahai, Cell communication networks in cancer invasion, *Curr. Opin. Cell Biol.* **23** (2011) 621–629.
10. M. A. J. Chaplain, M. Lachowicz, Z. Szymańska and D. Wrzosek, Mathematical modelling of cancer invasion: The importance of cell–cell adhesion and cell–matrix adhesion, *Math. Models Methods Appl. Sci.* **21** (2011) 719–743.
11. M. A. J. Chaplain and G. Lolas, Mathematical modelling of cancer invasion of tissue: Dynamic heterogeneity, *Netw. Heterog. Media* **1** (2006) 399–439.
12. A. Chapman, L. F. del Ama, J. Ferguson, J. Kamarashev, C. Wellbrock and A. Hurlstone, Heterogeneous tumor subpopulations cooperate to drive invasion, *Cell Rep.* **8** (2014) 688–695.
13. J. W. Cholewa and T. Dlotko, *Global Attractors in Abstract Parabolic Problems*, Vol. 278 (Cambridge Univ. Press, 2000).
14. C. Cillo, J. E. Dick, V. Ling and R. P. Hill, Generation of drug-resistant variants in metastatic B16 mouse melanoma cell lines, *Cancer Res.* **47** (1987) 2604–2608.
15. D. Cunningham and Z. You, *In vitro* and *in vivo* model systems used in prostate cancer research, *J. Biol. Meth.* **2** (2015) e17.
16. T. L. Davis, I. Rabinovitz, B. W. Futscher, M. Schnölzer, F. Burger, Y. Liu, M. Kulesz-Martin and A. E. Cress, Identification of a novel structural variant of the $\alpha 6$ integrin, *J. Biol. Chem.* **276** (2001) 26099–26106.
17. M. Delcommenne and C. H. Streuli, Control of integrin expression by extracellular matrix, *J. Biol. Chem.* **270** (1995) 26794–26801.
18. J. J. Deman, L. C. Vakaet and E. A. Bruyneel, Cell size and mutual cell adhesion. II. Evidence for a relation between cell size, long-range electrostatic repulsion and intercellular adhesiveness during density-regulated growth in suspension, *J. Membr. Biol.* **26** (1976) 205–215.
19. P. Domschke, D. Trucu, A. Gerisch and M. A. J. Chaplain, Mathematical modelling of cancer invasion: Implications of cell adhesion variability for tumour infiltrative growth patterns, *J. Theor. Biol.* **361** (2014) 41–60.
20. R. Eftimie, G. de Vries, M. A. Lewis and F. Lutscher, Modeling group formation and activity patterns in self-organizing collectives of individuals, *Bull. Math. Biol.* **69** (2007) 1537–1565.
21. L. C. Evans, *Partial Differential Equations*, Graduate Studies in Mathematics, Vol. 19 (Amer. Math. Soc., 2010).
22. J. Fan and K. Zhao, A note on a 3D haptotaxis model of cancer invasion, *Appl. Math. Res. Exp.* **2014** (2014) 74–86.
23. B. Geiger, Long-range morphogenetic signals and cell adhesion, *BioEssays* **13** (1991) 665–666.
24. A. Gerisch and M. A. J. Chaplain, Mathematical modelling of cancer cell invasion of tissue: Local and non-local models and the effect of adhesion, *J. Theor. Biol.* **250** (2008) 684–704.
25. J. E. F. Green, S. L. Waters, J. P. Whiteley, L. Edelstein-Keshet, K. M. Shakesheff and H. M. Byrne, Non-local models for the formation of hepatocyte-stellate cell aggregates, *J. Theor. Biol.* **267** (2010) 106–120.
26. D. Hanahan and R. A. Weinberg, The hallmarks of cancer, *Cell* **100** (2000) 57–70.
27. D. Henry, *Geometric Theory of Semilinear Parabolic Systems*, Vol. 840 (Springer-Verlag, 1981).
28. R. P. Hill, A. F. Chambers and V. Ling, Dynamic heterogeneity: Rapid generation of metastatic variants in mouse B16 melanoma cells, *Science* **224** (1984) 998–1001.

29. T. Hillen, C. Rohde and F. Lutscher, Existence of weak solutions for a hyperbolic model of chemosensitive movement, *J. Math. Anal. Appl.* **260** (2001) 173–199.
30. S. Huang and S. Chakrabarty, Regulation of fibronectin and laminin receptor expression, fibronectin and laminin secretion in human colon cancer cells by transforming growth factor- β 1, *Int. J. Cancer* **57** (1994) 742–746.
31. B. Kaminska, A. Wesolowska and M. Danilkiewicz, TGF beta signalling and its role in tumour pathogenesis, *Acta Biochim. Pol.* **52** (2005) 329.
32. E. Kang and J. Lee, Global solutions to chemotaxis–haptotaxis tumor invasion system with tissue re-establishment, *J. Chungcheong Math. Soc.* **28** (2015) 161–172.
33. L. Khalique, A. Ayhan, M. E. Weale, I. J. Jacobs, S. J. Ramus and S. A. Gayther, Genetic intra-tumour heterogeneity in epithelial ovarian cancer and its implications for molecular diagnosis of tumours, *J. Pathol.* **211** (2007) 286–295.
34. S. A. Khan, J. Joyce and T. Tsuda, Quantification of active and total transforming growth factor- β levels in serum and solid organ tissues by bioassay, *BMC Res. Notes* **5** (2012) 1.
35. Y. Kidera *et al.*, Reduction of lung metastasis, cell invasion and adhesion in mouse melanoma by statin-induced blockade of the rho/rho-associated coiled-coil-containing protein kinase pathway, *J. Exp. Clin. Cancer Res.* **29** (2010) 127.
36. Y. Kim and H. G. Othmer, A hybrid model of tumor-stromal interactions in breast cancer, *Bull. Math. Biol.* **75** (2013) 1304–1350.
37. A. K. Laird, Dynamics of tumor growth, *Brit. J. Cancer* **18** (1964) 490–502.
38. J. T. Leith, S. Michelson and A. S. Glicksman, Competitive exclusion of clonal subpopulations in heterogeneous tumours after stromal injury, *Brit. J. Cancer* **59** (1989) 22–27.
39. J. Liu, X. He, Y. Qi, X. Tian, S. J. Monkley, D. R. Critchley, S. A. Corbett, S. F. Lowry, A. M. Graham and S. Li, Talin1 regulates integrin turnover to promote embryonic epithelial morphogenesis, *Molec. Cell Biol.* **31** (2011) 3366–3377.
40. V. H. Lobert, A. Brech, N. M. Pedersen, J. Wesche, A. Oppelt, L. Malerød and H. Stenmark, Ubiquitination of α 5 β 1 integrin controls fibroblast migration through lysosomal degradation of fibronectin-integrin complexes, *Dev. Cell* **19** (2010) 148–159.
41. K. R. Loeb and L. A. Loeb, Significance of multiple mutations in cancer, *Carcinogenesis* **21** (2000) 379–385.
42. F. A. Mamuya and M. K. Duncan, α V integrins and TGF- β -induced EMT: A circle of regulation, *J. Cell Molec. Med.* **16** (2012) 445–455.
43. M. M. Mareel, P. De Baetselier and F. M. Van Roy, *Mechanisms of Invasion and Metastasis* (CRC Press, 1991).
44. S. Markowitz *et al.*, Inactivation of the type a TGF- β receptor in colon cancer cells with microsatellite instability, *Science* **268** (1995) 1336–1338.
45. A. Marusyk and K. Polyak, Tumor heterogeneity: Causes and consequences, *Biochim. Biophys. Acta* **1805** (2010) 105.
46. S. Michelson and J. Leith, Autocrine and paracrine growth factors in tumor growth: A mathematical model, *Bull. Math. Biol.* **53** (1991) 639–656.
47. K. Miyazono, Transforming growth factor- β signaling in epithelial–mesenchymal transition and progression of cancer, *P. Jpn. Acad. B-Phys.* **85** (2009) 314–323.
48. A. Mogilner and L. Edelstein-Keshet, Selecting a common direction: I. How orientational order can arise from simple contact responses between interacting cells, *J. Math. Biol.* **33** (1995) 619–660.
49. A. Mogilner, L. Edelstein-Keshet and G. B. Ermentrout, Selecting a common direction: II. Peak-like solutions representing total alignment of cell clusters, *J. Math. Biol.* **34** (1996) 811–842.

50. F. Morani, S. Phadngam, C. Follo, R. Titone, V. Thongrakard, A. Galetto, O. Alabiso and C. Isidoro, PTEN deficiency and mutant p53 confer glucose-addiction to thyroid cancer cells: Impact of glucose depletion on cell proliferation, cell survival, autophagy and cell migration, *Genes Cancer* **5** (2014) 226–239.
51. A. Moustakas and C.-H. Heldin, Signaling networks guiding epithelial–mesenchymal transitions during embryogenesis and cancer progression, *Cancer Sci.* **98** (2007) 1512–1520.
52. A. Nawshad, D. LaGamba, A. Polad and E. D. Hay, Transforming growth factor- β signaling during epithelial–mesenchymal transformation: Implications for embryogenesis and tumor metastasis, *Cells Tissues Organs* **179** (2005) 11–23.
53. H. Nessyahu and E. Tadmor, Non-oscillatory central differencing for hyperbolic conservation laws, *J. Comput. Phys.* **87** (1990) 408–463.
54. G. L. Nicholson, Tumor cell instability, diversification and progression to the metastatic phenotype: From oncogene to oncofetal expression, *Cancer Res.* **47** (1987) 1473–1487.
55. N. Outada, N. Vauchelet, T. Akrid and M. Khaladi, From kinetic theory of multicellular systems to hyperbolic tissue equations: Asymptotic limits and computing, *Math. Models Methods Appl. Sci.* **26** (2016) 2709–2734.
56. K. J. Painter, N. J. Armstrong and J. A. Sherratt, The impact of adhesion on cellular invasion processes in cancer and development, *J. Theor. Biol.* **264** (2010) 1057–1067.
57. D. R. Principe, J. A. Doll, J. Bauer, B. Jung, H. G. Munshi, L. Bartholin, B. Pasche, C. Lee and P. J. Grippo, TGF- β : Duality of function between tumor prevention and carcinogenesis, *J. Natl. Cancer I.* **106** (2014) djt369.
58. T. Runst and W. Sickel, *Sobolev Spaces of Fractional Order, Nemytskij Operators, and Nonlinear Partial Differential Equations*, Vol. 3 (Walter de Gruyter, 1996).
59. J. A. Sherratt, S. A. Gourley, N. J. Armstrong and K. J. Painter, Boundedness of solutions of a non-local reaction–diffusion model for adhesion in cell aggregation and cancer invasion, *Eur. J. Appl. Math.* **20** (2009) 123–144.
60. Z. Szymańska, C. M. Rodrigo, M. Lachowicz and M. A. J. Chaplain, Mathematical modelling of cancer invasion of tissue: The role and effect of nonlocal interactions, *Math. Models Methods Appl. Sci.* **19** (2009) 257–281.
61. Y. Tao, Global existence for a haptotaxis model of cancer invasion with tissue remodeling, *Nonlinear Anal. Real World Appl.* **12** (2011) 418–435.
62. Y. Tao and M. Winkler, Energy-type estimates and global solvability in a two-dimensional chemotaxis–haptotaxis model with remodeling of non-diffusible attractant, *J. Differential Equations* **257** (2014) 784–815.
63. S. Turner, J. A. Sherratt and D. Cameron, Tamoxifen treatment failure in cancer and the nonlinear dynamics of TGF β , *J. Theor. Biol.* **229** (2004) 101–111.
64. A. Van Schepdael, A. Carlier and L. Geris, *Sensitivity Analysis by Design of Experiments* (Springer, 2016).
65. L. Venkatraman, S.-M. Chia, B. C. Narmada, J. K. White, S. S. Bhowmick, C. F. Dewey, P. T. So, L. Tucker-Kellogg and H. Yu, Plasmin triggers a switch-like decrease in thrombospondin-dependent activation of TGF- β 1, *Biophys. J.* **103** (2012) 1060–1068.
66. H. Wang, V. Radjendirane, K. K. Wary and S. Chakrabarty, Transforming growth factor β regulates cell–cell adhesion through extracellular matrix remodeling and activation of focal adhesion kinase in human colon carcinoma moser cells, *Oncogene* **23** (2004) 5558–5561.

67. S. E. Wang, P. Hinow, N. Bryce, A. M. Weaver, L. Estrada, C. L. Arteaga and G. F. Webb, A mathematical model quantifies proliferation and motility effects of TGF- β on cancer cells, *Comput. Math. Methods Med.* **10** (2009) 71–83.
68. J. B. Weitzman, A. Chen and M. E. Hemler, Investigation of the role of $\beta 1$ integrins in cell–cell adhesion, *J. Cell Sci.* **108** (1995) 3635–3644.
69. M.-S. Wu, J.-T. Lin, P.-N. Hsu, C.-Y. Lin, Y.-T. Hsieh, Y.-H. Chiu, P.-R. Hsueh and K.-W. Liao, Preferential induction of transforming growth factor- β production in gastric epithelial cells and monocytes by *Helicobacter pylori* soluble proteins, *J. Infect. Diseases* **196** (2007) 1386–1393.